

## APCluster

### An R Package for Affinity Propagation Clustering

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## Scope and Purpose of this Document

This document is a user manual for the R package `apcluster` [1]. It is only meant as a gentle introduction into how to use the basic functions implemented in this package. Not all features of the R package are described in full detail. Such details can be obtained from the documentation enclosed in the R package. Further note the following: (1) this is neither an introduction to affinity propagation nor to clustering in general; (2) this is not an introduction to R. If you lack the background for understanding this manual, you first have to read introductory literature on these subjects.

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## 1 Introduction

Affinity propagation (AP) is a relatively new clustering algorithm that has been introduced by Brendan J. Frey and Delbert Dueck [5].<sup>1</sup> The authors themselves describe affinity propagation as follows:<sup>2</sup>

*“An algorithm that identifies exemplars among data points and forms clusters of data points around these exemplars. It operates by simultaneously considering all data point as potential exemplars and exchanging messages between data points until a good set of exemplars and clusters emerges.”*

AP has been applied in various fields recently, among which bioinformatics is becoming increasingly important. Frey and Dueck have made their algorithm available as Matlab code.<sup>1</sup> Matlab, however, is relatively uncommon in bioinformatics. Instead, the statistical computing platform R has become a widely accepted standard in this field. In order to leverage affinity propagation for bioinformatics applications, we have implemented affinity propagation as an R package. Note, however, that the given package is in no way restricted to bioinformatics applications. It is as generally applicable as Frey’s and Dueck’s original Matlab code.<sup>1</sup>

Starting with Version 1.1.0, the `apcluster` package also features exemplar-based agglomerative clustering which can be used as a clustering method on its own or for creating a hierarchy of clusters that have been computed previously by affinity propagation.

## 2 Installation

### 2.1 Installation via CRAN

The R package `apcluster` (current version: 1.2.0) is part of the *Comprehensive R Archive Network (CRAN)*<sup>3</sup>. The simplest way to install the package, therefore, is to enter the following command into your R session:

```
> install.packages("apcluster")
```

### 2.2 Manual installation

If, for what reason ever, you prefer to install the package manually, download the package file suitable for your computer system and copy it to your harddisk. Open the package’s page at CRAN<sup>4</sup> and the proceed as follows.

---

<sup>1</sup><http://www.psi.toronto.edu/affinitypropagation/>

<sup>2</sup>quoted from <http://www.psi.toronto.edu/affinitypropagation/faq.html#def>

<sup>3</sup><http://cran.r-project.org/>

<sup>4</sup><http://cran.r-project.org/web/packages/apcluster/index.html>

### Manual installation under Windows

1. Download `apcluster_1.2.0.zip` and save it to your harddisk
2. Open the R GUI and select the menu entry

`Packages | Install package(s) from local zip files...`

(if you use R in a different language, search for the analogous menu entry). In the file dialog that opens, go to the folder where you placed `apcluster_1.2.0.zip` and select this file. The package should be installed now.

### Manual installation under Linux/UNIX/MacOS

1. Download `apcluster_1.2.0.tar.gz` and save it to your harddisk.
2. Open a shell window and change to the directory where you put `apcluster_1.2.0.tar.gz`. Enter

`R CMD INSTALL apcluster_1.2.0.tar.gz`

to install the package.

## 2.3 Compatibility issues

Both the Windows and the Linux/UNIX/MacOS version available from CRAN have been built using the latest version, R 2.14.0. However, the package should work without severe problems on R versions  $\geq 2.10.1$ .

## 3 Getting Started

To load the package, enter the following in your R session:

```
> library(apcluster)
```

If this command terminates without any error message or warning, you can be sure that the package has been installed successfully. If so, the package is ready for use now and you can start clustering your data with affinity propagation.

The package includes both a user manual (this document) and a reference manual (help pages for each function). To view the user manual, enter

```
> vignette("apcluster")
```

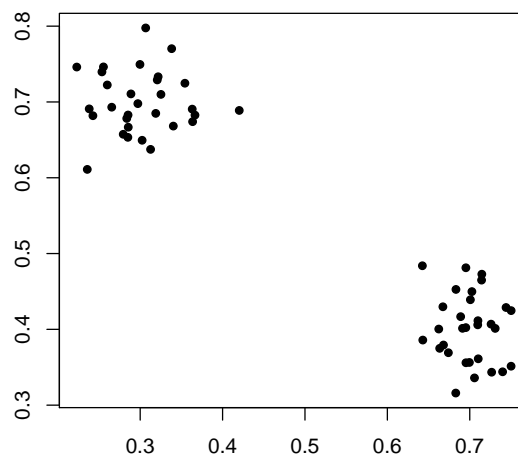
Help pages can be viewed using the `help` command. It is recommended to start with

```
> help(apcluster)
```

Affinity propagation does not require the data samples to be of any specific kind or structure. AP only requires a *similarity matrix*, i.e., given  $l$  data samples, this is an  $l \times l$  real-valued matrix  $S$ , in which an entry  $S_{ij}$  corresponds to a value measuring how similar sample  $i$  is to sample  $j$ . AP does not require these values to be in a specific range. Values can be positive or negative. AP does not even require the similarity matrix to be symmetric (although, in most applications, it will be symmetric anyway). A value of  $-\infty$  is interpreted as “absolute dissimilarity”. The higher a value, the more similar two samples are considered.

To get a first impression, let us create a random data set in  $\mathbb{R}^2$  as the union of two “Gaussian clouds”:

```
> cl1 <- cbind(rnorm(30, 0.3, 0.05), rnorm(30, 0.7, 0.04))
> cl2 <- cbind(rnorm(30, 0.7, 0.04), rnorm(30, 0.4, .05))
> x1 <- rbind(cl1, cl2)
> plot(x1, xlab="", ylab="", pch=19, cex=0.8)
```



Now we have to create a similarity matrix. The package `apcluster` offers several different ways for doing that (see Section 7 below). Let us start with the default similarity measure used in the papers of Frey and Dueck — negative squared distances:

```
> s1 <- negDistMat(x1, r=2)
```

We are now ready to run affinity propagation:

```
> apres1a <- apcluster(s1)
```

The function `apcluster()` creates an object belonging to the S4 class `APResult` which is defined by the present package. To get detailed information on which data are stored in such objects, enter

```
> help(APResult)
```

The simplest thing we can do is to enter the name of the object (which implicitly calls `show()`) to get a summary of the clustering result:

```
> apres1a
```

APResult object

```
Number of samples      = 60
Number of iterations   = 131
Input preference       = -0.1416022
Sum of similarities    = -0.1955119
Sum of preferences     = -0.2832044
Net similarity         = -0.4787163
Number of clusters     = 2
```

Exemplars:

```
25 44
```

Clusters:

Cluster 1, exemplar 25:

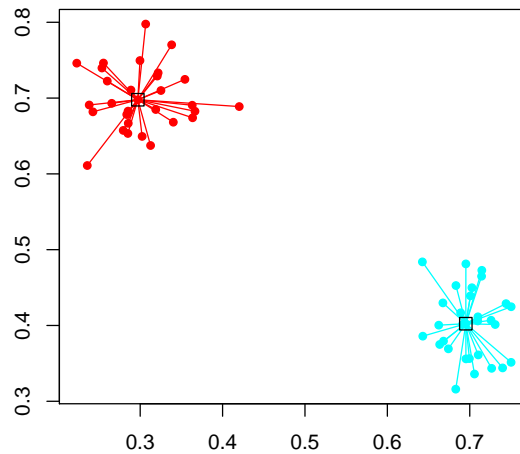
```
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26
27 28 29 30
```

Cluster 2, exemplar 44:

```
31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53
54 55 56 57 58 59 60
```

For two-dimensional data sets, the `apcluster` package allows for plotting the original data set along with a clustering result:

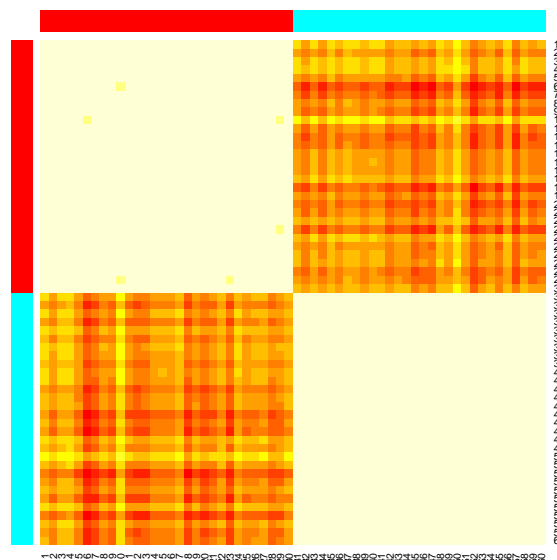
```
> plot(apres1a, x1)
```



In this plot, each color corresponds to one cluster. The exemplar of each cluster is marked by a box and all cluster members are connected to their exemplars with lines.

If `plot()` is called with the similarity matrix as second argument, a heatmap is created:

```
> plot(apres1a, s1)
```



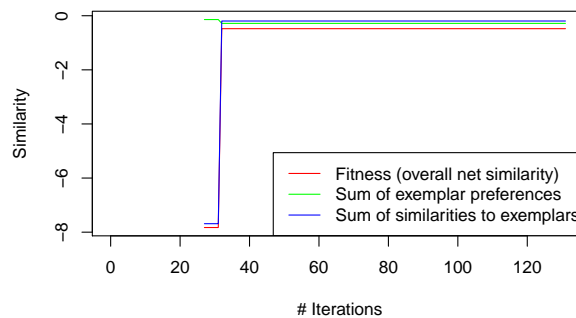
In the heatmap, the samples are grouped according to clusters. The above heatmap confirms again that there are two main clusters in the data.

Suppose we want to have better insight into what the algorithm did in each iteration. For this purpose, we can supply the option `details=TRUE` to `apcluster()`:

```
> apres1b <- apcluster(s1, details=TRUE)
```

This option tells the algorithm to keep a detailed log about its progress. For example, this allows us to plot the three performance measures that AP uses internally for each iteration:

```
> plot(apres1b)
```



These performance measures are:

1. Sum of exemplar preferences
2. Sum of similarities of exemplars to their cluster members
3. Net fitness: sum of the two former

For details, the user is referred to the original affinity propagation paper [5] and the supplementary material published on the affinity propagation Web page.<sup>1</sup> We see from the above plot that the algorithm has not made any change for the last 100 (of 131!) iterations. AP, through its parameter `convits`, allows to control for how long AP waits for a change until it terminates (the default is `convits=100`). If the user has the feeling that AP will probably converge quicker on his/her data set, a lower value can be used:

```
> apres1c <- apcluster(s1, convits=15, details=TRUE)
> apres1c
```

APResult object

```
Number of samples      = 60
Number of iterations   = 46
Input preference       = -0.1416022
Sum of similarities    = -0.1955119
Sum of preferences     = -0.2832044
Net similarity         = -0.4787163
Number of clusters     = 2
```



Exemplars:

25 44

Clusters:

Cluster 1, exemplar 25:

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26  
27 28 29 30

Cluster 2, exemplar 44:

31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53  
54 55 56 57 58 59 60

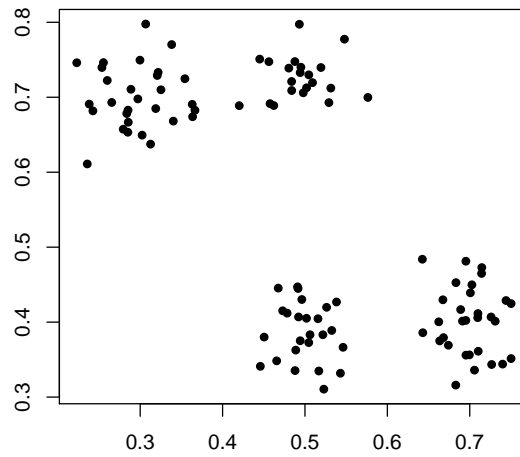
## 4 Adjusting Input Preferences

Apart from the similarity itself, the most important input parameter of AP is the so-called *input preference* which can be interpreted as the tendency of a data sample to become an exemplar (see [5] and supplementary material on the AP homepage<sup>1</sup> for a more detailed explanation). This input preference can either be chosen individually for each data sample or it can be a single value shared among all data samples. Input preferences largely determine the number of clusters, in other words, how fine- or coarse-grained the clustering result will be.

The input preferences one can specify for AP are roughly in the same range as the similarity values, but they do not have a straightforward interpretation. Frey and Dueck have introduced the following rule of thumb: “*The shared value could be the median of the input similarities (resulting in a moderate number of clusters) or their minimum (resulting in a small number of clusters).*” [5]

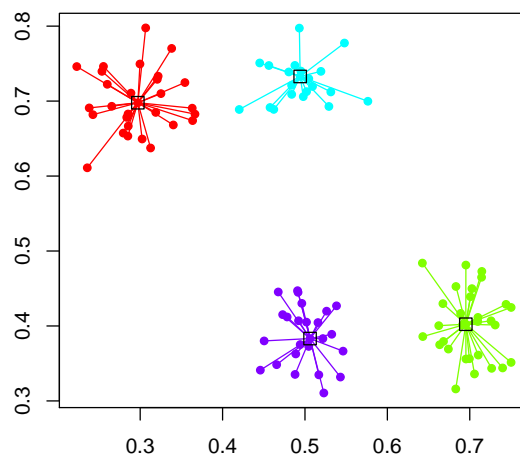
Our AP implementation uses the median rule by default if the user does not supply a custom value for the input preferences. In order to provide the user with a knob that is — at least to some extent — interpretable, the function `apcluster` has a new argument `q` that allows to set the input preference to a certain quantile of the input similarities: resulting in the median for `q=0.5` and in the minimum for `q=0`. As an example, let us add two more “clouds” to the data set from above:

```
> c13 <- cbind(rnorm(20, 0.50, 0.03), rnorm(20, 0.72, 0.03))
> c14 <- cbind(rnorm(25, 0.50, 0.03), rnorm(25, 0.42, 0.04))
> x2 <- rbind(x1, c13, c14)
> s2 <- negDistMat(x2, r=2)
> plot(x2, xlab="", ylab="", pch=19, cex=0.8)
```



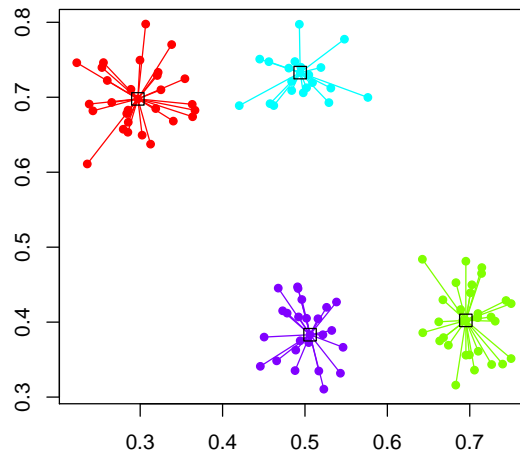
For the default setting, we obtain the following result:

```
> apres2a <- apcluster(s2)
> plot(apres2a, x2)
```



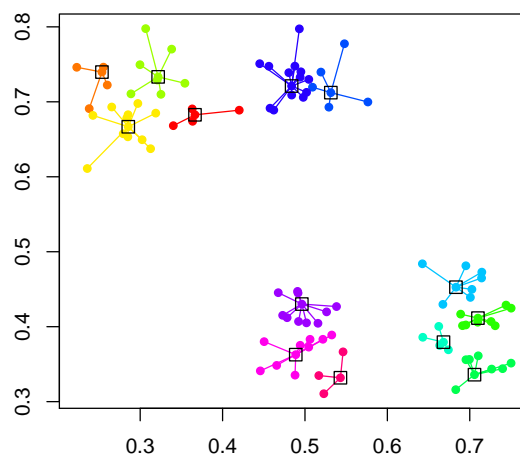
For the minimum of input similarities, we obtain the following result:

```
> apres2b <- apcluster(s2, q=0)
> plot(apres2b, x2)
```



So we see that AP is quite robust against a reduction of input preferences in this example which may be caused by the clear separation of the four clusters. If we increase input preferences, however, we can force AP to split the four clusters into smaller sub-clusters:

```
> apres2c <- apcluster(s2, q=0.8)
> plot(apres2c, x2)
```



Note that the input preference used by AP can be recovered from the output object (no matter which method to adjust input preferences has been used). On the one hand, the value is printed if

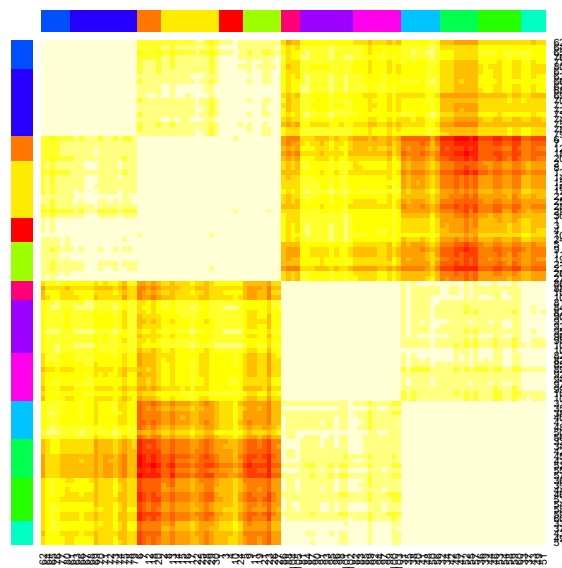
the object is displayed (by `show` or by entering the output object's name). On the other hand, the value can be accessed directly via the slot `p`:

```
> apres2c@p

[1] -0.008941595
```

As noted above already, we can produce a heatmap by calling `plot()` with an `APResult` object as first and the corresponding similarity matrix as second argument:

```
> plot(apres2c, s2)
```



The order in which the clusters are arranged in the heatmap is determined by means of joining the cluster agglomeratively (see Section 5 below). Although the affinity propagation result contains 13 clusters, the heatmap indicates that there are actually four clusters which can be seen as very brightly colored squares along the diagonal. We also see that there seem to be two pairs of adjacent clusters, which can be seen from the fact that there are two relatively light-colored blocks along the diagonal encompassing two of the four clusters in each case. If we look back at how the data have been created (see also plots above), this is exactly what is to be expected.

The above example with  $q=0$  demonstrates that setting input preferences to the minimum of input similarities does not necessarily result in a very small number of clusters (like one or two). This is due to the fact that input preferences need not necessarily be exactly in the range of the similarities. To determine a meaningful range, an auxiliary function is available which, in line with Frey's and Dueck's Matlab code,<sup>1</sup> allows to compute a minimum value (for which one or at most two clusters would be obtained) and a maximum value (for which as many clusters as data samples would be obtained):

```
> preferenceRange(s2)
```

```
[1] -6.271275e+00 -5.292318e-06
```

The function returns a two-element vector with the minimum value as first and the maximum value as second entry. The computations are done approximately by default. If one is interested in exact bounds, supply `exact=TRUE` (resulting in longer computation times).

Many clustering algorithms need to know a pre-defined number of clusters. This is often a major nuisance, since the exact number of clusters is hard to know for non-trivial (in particular, high-dimensional) data sets. AP avoids this problem. If, however, one still wants to require a fixed number of clusters, this has to be accomplished by a search algorithm that adjusts input preferences in order to produce the desired number of clusters in the end. For convenience, this search algorithm is available as a function `apclusterK()` (analogous to Frey's and Dueck's Matlab implementation<sup>1</sup>). We can use this function to force AP to produce only two clusters (merging the two pairs of adjacent clouds into one cluster each):

```
> apres2d <- apclusterK(s2, K=2, verbose=TRUE)
```

```
Trying p = -0.006276562
```

```
Number of clusters: 16
```

```
Trying p = -0.06271799
```

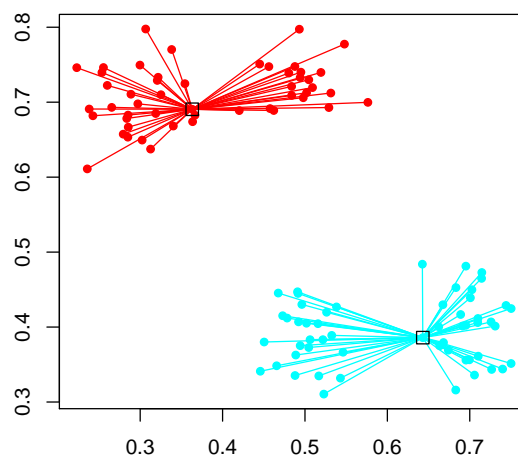
```
Number of clusters: 4
```

```
Trying p = -0.6271322
```

```
Number of clusters: 2
```

```
Number of clusters: 2 for p = -0.6271322
```

```
> plot(apres2d, x2)
```



## 5 Exemplar-based Agglomerative Clustering

The function `aggExCluster()` realizes what can best be described as “exemplar-based agglomerative clustering”, i.e. agglomerative clustering whose merging objective is geared towards the identification of meaningful exemplars. `aggExCluster()` only requires a matrix of pairwise similarities.

### 5.1 Getting started

Let us start with a simple example:

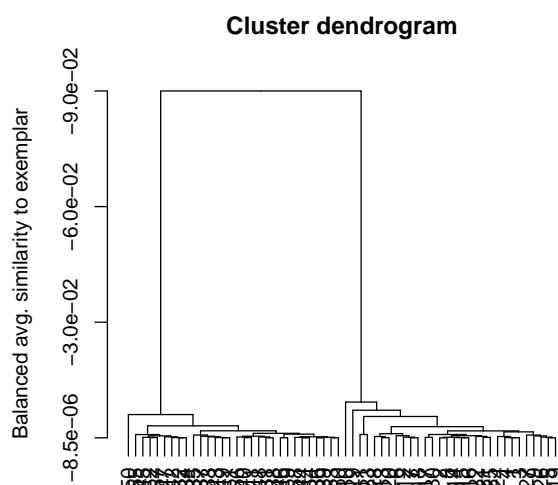
```
> aggres1a <- aggExCluster(s1)
> aggres1a
```

AggExResult object

```
Number of samples      = 60
Maximum number of clusters = 60
```

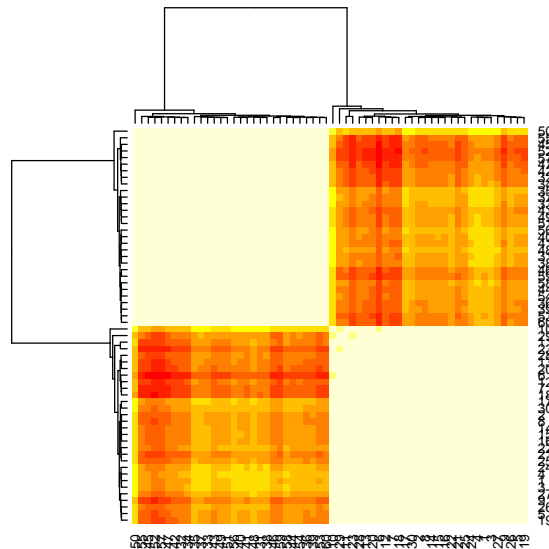
The output object `aggres1a` contains the complete cluster hierarchy. As obvious from the above example, the `show()` method only displays the most basic information. Calling `plot()` on an object that was the result of `aggExCluster()` (an object of class `AggExResult`), a dendrogram is plotted:

```
> plot(aggres1a)
```



The heights of the merges in the dendrogram correspond to the merging objective: the higher the vertical bar of a merge, the less similar the two clusters have been. The dendrogram, therefore, clearly indicates two clusters. Calling `plot()` for an `AggExResult` object along with the similarity matrix as second argument produces a heatmap, where the dendrogram is plotted on top and to the left of the heatmap:

```
> plot(aggres1a, s1)
```



Once we have confirmed the number of clusters, which is clearly 2 according to the dendrogram and the heatmap above, we can extract the level with two clusters from the cluster hierarchy. In concordance with standard R terminology, the function for doing this is called `cutree()`:

```
> cl1a <- cutree(aggres1a, k=2)
> cl1a
```

ExClust object

```
Number of samples = 60
Number of clusters = 2
```

Exemplars:

```
44 25
```

Clusters:

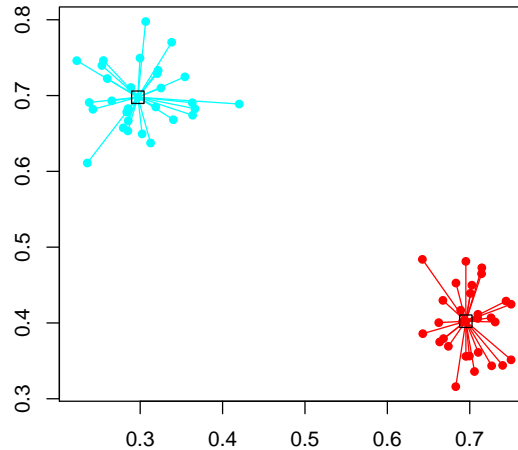
Cluster 1, exemplar 44:

```
50 55 45 52 57 47 42 32 34 35 37 33 43 49 51 56 40 41 48 31 38 46 59
58 44 54 36 39 53 60
```

Cluster 2, exemplar 25:

```
10 29 11 23 28 13 20 6 12 7 18 17 30 2 8 14 15 16 22 21 25 24 4 1 3
27 9 26 5 19
```

```
> plot(cl1a, x1)
```



## 5.2 Merging clusters obtained from affinity propagation

The most important application of `aggExCluster()` (and the reason why it is part of the `apcluster` package) is that it can be used for creating a hierarchy of clusters starting from a set of clusters previously computed by affinity propagation. The examples in Section 4 indicate that it may sometimes be tricky to define the right input preference. Exemplar-based agglomerative clustering on affinity propagation results provides an additional tool for finding the right number of clusters.

Let us revisit the four-cluster example from Section 4. We can apply `aggExCluster()` to an affinity propagation result if we run it on the same similarity matrix (as first argument) and supply the affinity propagation result as second argument:

```
> aggres2a <- aggExCluster(s2, apres2c)
> aggres2a
```

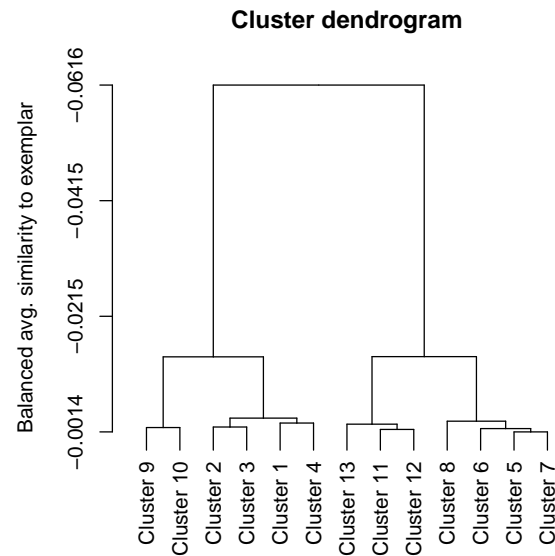
AggExResult object

```
Number of samples      = 105
Maximum number of clusters = 13
```

The result `apres2c` had 13 clusters. `aggExCluster()` successively joins these clusters until only one cluster is left. The dendrogram of this cluster hierarchy is given as follows:

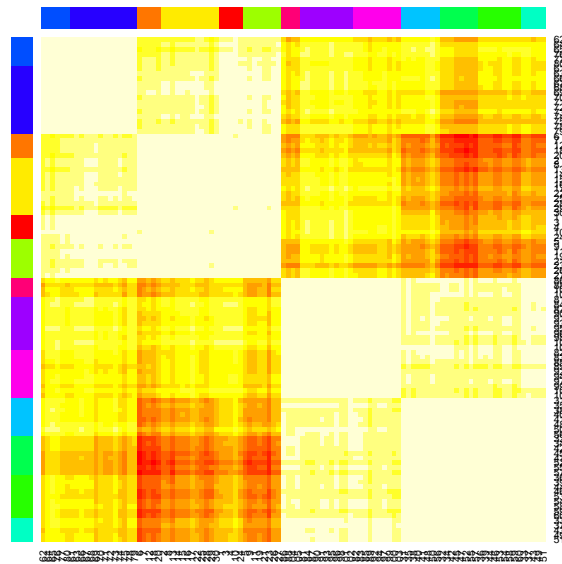
```
> plot(aggres2a)
```





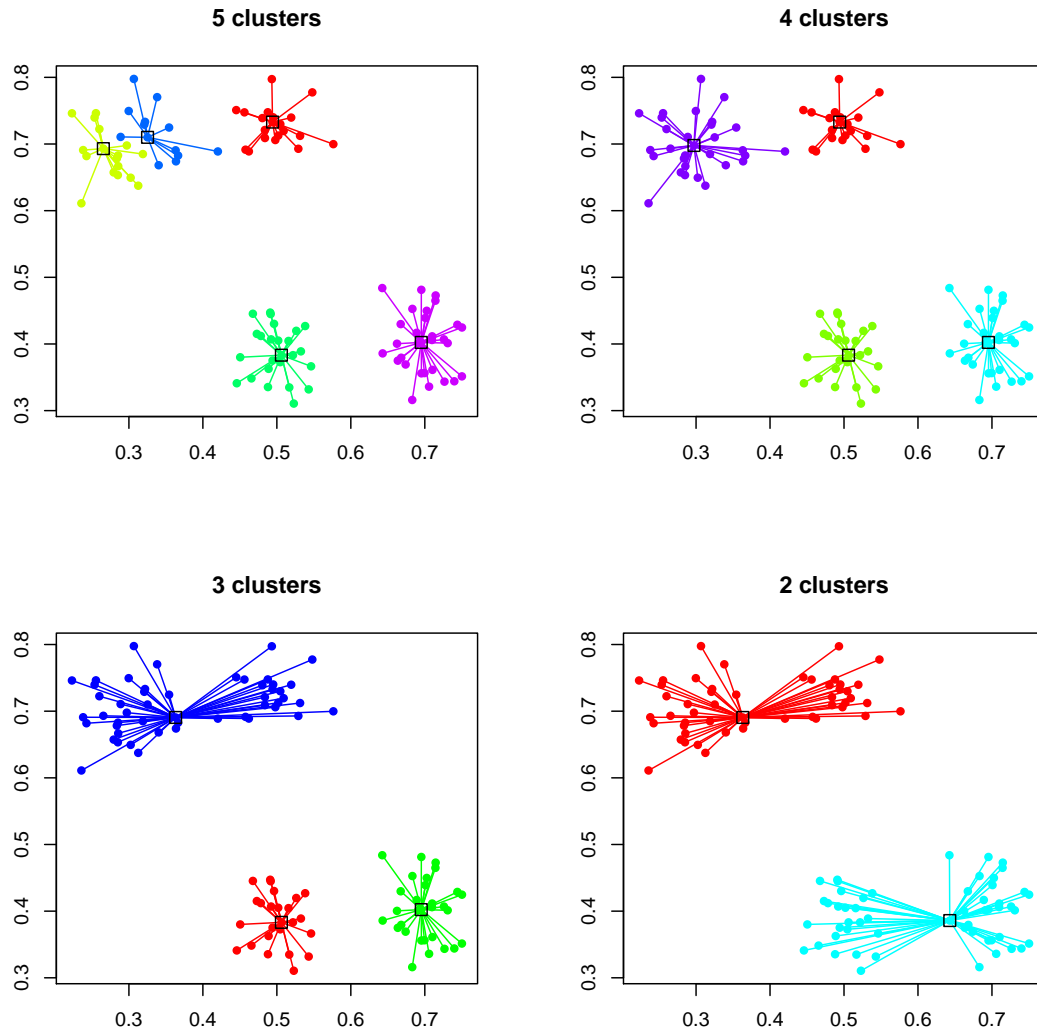
The following heatmap coincides with the one shown in Section 4 above. This is not surprising, since the heatmap plot for an affinity propagation result uses `aggExCluster()` internally to arrange the clusters:

```
> plot(aggres2a, s2)
```



Once we are more or less sure about the number of clusters, we extract the right clustering level from the hierarchy. For demonstration purposes, we do this for  $k = 5, \dots, 2$  in the following plots:

```
> par(mfrow=c(2,2))
> for (k in 5:2)
+   plot(aggres2a, x2, k=k, main=paste(k, "clusters"))
```



There is one obvious, but important, condition: applying `aggExCluster()` to an affinity propagation result only makes sense if the number of clusters to start from is at least as large as the number of true clusters in the data set. Clearly, if the number of clusters is already too small, then merging will make the situation only worse.

### 5.3 Details on the merging objective

Like any other agglomerative clustering method (see, e.g., [6, 10, 12]), `aggExCluster()` merges clusters until only one cluster containing all samples is obtained. In each step, two clusters

are merged into one, i.e. the number of clusters is reduced by one. The only aspect in which `aggExCluster()` differs from other methods is the merging objective.

Suppose we consider two clusters for possible merging, each of which is given by an index set:

$$I = \{i_1, \dots, i_{n_I}\} \text{ and } J = \{j_1, \dots, j_{n_J}\}$$

Then we first determine the potential *joint exemplar*  $\text{ex}(I, J)$  as the sample that maximizes the average similarity to all samples in the joint cluster  $I \cup J$ :

$$\text{ex}(I, J) = \underset{i \in I \cup J}{\operatorname{argmax}} \frac{1}{n_I + n_J} \cdot \sum_{j \in I \cup J} S_{ij}$$

Recall that  $\mathbf{S}$  denotes the similarity matrix and  $S_{ij}$  corresponds to the similarity of the  $i$ -th and the  $j$ -th sample. Then the merging objective is computed as

$$\text{obj}(I, J) = \frac{1}{2} \cdot \left( \frac{1}{n_I} \cdot \sum_{j \in I} S_{\text{ex}(I, J)j} + \frac{1}{n_J} \cdot \sum_{k \in J} S_{\text{ex}(I, J)k} \right),$$

which can be best described as “*balanced average similarity to the joint exemplar*”. In each step, `aggExCluster()` considers all pairs of clusters in the current cluster set and joins that pair of clusters whose merging objective is maximal. The rationale behind the merging objective is that those two clusters should be joined that are best described by a joint exemplar.

## 6 A Toy Example with Biological Sequences

As noted in the introduction above, one of the goals of this package is to leverage affinity propagation in bioinformatics applications. In order to demonstrate the usage of the package in a biological application, we consider a small toy example here.

The package comes with a toy data set `ch22Promoters` that consists of promoter regions of 150 random genes from the human chromosome no. 22 (according to the human genome assembly hg18). Each sequence consists of the 1000 bases upstream of the transcription start site of each gene. Suppose we want to cluster these sequences in order to find out whether groups of promoters can be identified on the basis of the sequence only and, if so, to identify exemplars that are most typical for these groups.

```
> data(ch22Promoters)
> names(ch22Promoters)[1:5]

[1] "NM_001169111" "NM_012324"      "NM_144704"      "NM_002473"
[5] "NM_001184970"

> substr(ch22Promoters[1:5], 951, 1000)

NM_001169111
"GCACGCGCTGAGAGCCTGTCAGCGGCTGCGCCCGTGTGCGCATGCGCAGC"
```

```

NM_012324
"CCGCCTCCCCCGCCGCCCTCCCCGCGCCGCCGCGGAGTCCGGGCGAGGTG"
NM_144704
"GTGCTGGGCCCCGCGGGCTCCCCGCGCCGAGTGCAAACGCAGCGCCAGACA"
NM_002473
"CAGGCTCCGCCCCGGAGCCGGCTCCCGGCTGGGAATGGTCCCGCGGCTCC"
NM_001184970
"GGGGCGGGGCTCGGTGTCCGGTAGCCAATGGACAGAGCCCAGCGGGAGCG"

```

Obviously, these are classical nucleotide sequences, each of which is identified by the RefSeq identifier of the gene the promoter sequence stems from.

In order to compute a similarity matrix for this data set, we choose (without further justification, just for demonstration purposes) the simple *spectrum kernel* [8] with a sub-sequence length of  $k = 6$ . We use the implementation from the *kernlab* package [7] to compute the similarity matrix in a convenient way:

```

> library(kernlab)
> promSim <- kernelMatrix(stringdot(length=6, type="spectrum"), ch22Promoters)
> rownames(promSim) <- names(ch22Promoters)
> colnames(promSim) <- names(ch22Promoters)

```

Now we run affinity propagation on this similarity matrix:

```

> promAP <- apcluster(promSim, q=0)
> promAP

```

APResult object

```

Number of samples      = 150
Number of iterations   = 185
Input preference       = 0.01367387
Sum of similarities    = 57.76715
Sum of preferences     = 0.109391
Net similarity         = 57.87655
Number of clusters     = 8

```

Exemplars:

```

NM_001199580 NM_022141 NM_152868 NM_001128633 NM_052945 NM_001099294
NM_152513 NM_080764

```

Clusters:

```

Cluster 1, exemplar NM_001199580:
NM_001169111 NM_012324 NM_144704 NM_002473 NM_005198 NM_004737
NM_007194 NM_014292 NM_001199580 NM_032758 NM_003325 NM_014876
NM_000407 NM_053004 NM_023004 NM_001130517 NM_002882 NM_001169110
NM_020831 NM_001195071 NM_031937 NM_001164502 NM_152299 NM_014509
NM_138433 NM_006487 NM_005984 NM_001085427 NM_013236

```

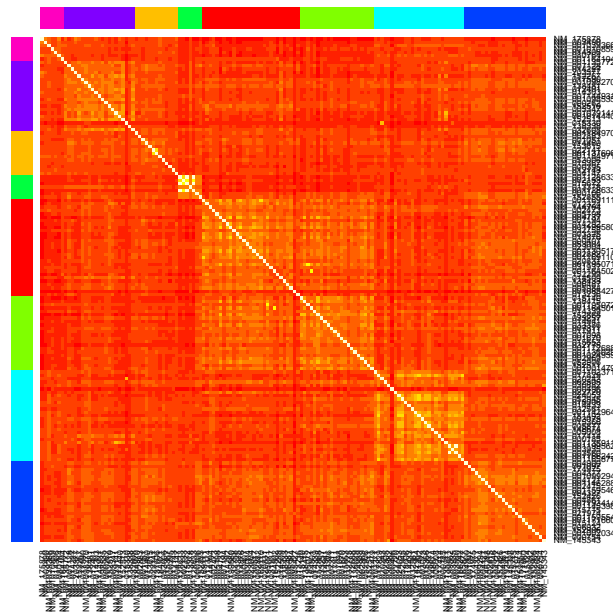
```

Cluster 2, exemplar NM_022141:
  NM_001184970 NM_002883 NM_001051 NM_014460 NM_153615 NM_022141
  NM_001137606 NM_001184971 NM_053006 NM_015367 NM_000395 NM_012143
  NM_004147
Cluster 3, exemplar NM_152868:
  NM_015140 NM_138415 NM_001195072 NM_001164501 NM_014339 NM_152868
  NM_033257 NM_014941 NM_033386 NM_007311 NM_017911 NM_007098
  NM_001670 NM_015653 NM_032775 NM_001172688 NM_001136029 NM_001024939
  NM_002969 NM_052906 NM_152511 NM_001001479
Cluster 4, exemplar NM_001128633:
  NM_001128633 NM_002133 NM_015672 NM_013378 NM_001128633 NM_000106
  NM_152855
Cluster 5, exemplar NM_052945:
  NM_001102371 NM_017829 NM_020243 NM_002305 NM_030882 NM_000496
  NM_022720 NM_024053 NM_052945 NM_018006 NM_014433 NM_032561
  NM_001142964 NM_181492 NM_003073 NM_015366 NM_005877 NM_148674
  NM_005008 NM_017414 NM_000185 NM_001135911 NM_001199562 NM_003935
  NM_003560 NM_001166242 NM_001165877
Cluster 6, exemplar NM_001099294:
  NM_004900 NM_001097 NM_174975 NM_004377 NM_001099294 NM_004121
  NM_001146288 NM_002415 NM_001159546 NM_004327 NM_152426 NM_004861
  NM_001193414 NM_001145398 NM_015715 NM_021974 NM_001159554
  NM_001171660 NM_015124 NM_006932 NM_152906 NM_001002034 NM_000754
  NM_145343
Cluster 7, exemplar NM_152513:
  NM_001135772 NM_007128 NM_014227 NM_203377 NM_152267 NM_017590
  NM_001098270 NM_138481 NM_012401 NM_014303 NM_001144931 NM_001098535
  NM_000262 NM_152510 NM_003347 NM_001039141 NM_001014440 NM_152513
  NM_015330 NM_138338 NM_032608
Cluster 8, exemplar NM_080764:
  NM_175878 NM_003490 NM_001039366 NM_001010859 NM_014306 NM_080764
  NM_001164104

```

So we obtain 8 clusters in total. The corresponding heatmap looks as follows:

```
> plot(promAP, promSim)
```

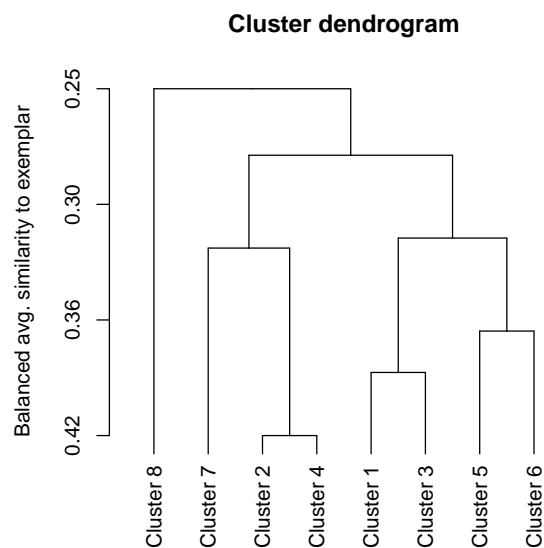


Let us now run agglomerative clustering to further join clusters.

```
> promAgg <- aggExCluster(promSim, promAP)
```

The resulting dendrogram is given as follows:

```
> plot(promAgg)
```



The dendrogram does not give a very clear indication about the best number of clusters. Let us adopt the viewpoint for a moment that 5 clusters are reasonable.

```
> prom5 <- cutree(promAgg, k=5)
> prom5
```

ExClust object

```
Number of samples = 150
Number of clusters = 5
```

Exemplars:

```
NM_152513 NM_080764 NM_001128633 NM_001199580 NM_052945
```

Clusters:

Cluster 1, exemplar NM\_152513:

```
NM_001135772 NM_007128 NM_014227 NM_203377 NM_152267 NM_017590
NM_001098270 NM_138481 NM_012401 NM_014303 NM_001144931 NM_001098535
NM_000262 NM_152510 NM_003347 NM_001039141 NM_001014440 NM_152513
NM_015330 NM_138338 NM_032608
```

Cluster 2, exemplar NM\_080764:

```
NM_175878 NM_003490 NM_001039366 NM_001010859 NM_014306 NM_080764
NM_001164104
```

Cluster 3, exemplar NM\_001128633:

```
NM_001184970 NM_002883 NM_001051 NM_014460 NM_153615 NM_022141
NM_001137606 NM_001184971 NM_053006 NM_015367 NM_000395 NM_012143
NM_004147 NM_001128633 NM_002133 NM_015672 NM_013378 NM_001128633
NM_000106 NM_152855
```

Cluster 4, exemplar NM\_001199580:

```
NM_001169111 NM_012324 NM_144704 NM_002473 NM_005198 NM_004737
NM_007194 NM_014292 NM_001199580 NM_032758 NM_003325 NM_014876
NM_000407 NM_053004 NM_023004 NM_001130517 NM_002882 NM_001169110
NM_020831 NM_001195071 NM_031937 NM_001164502 NM_152299 NM_014509
NM_138433 NM_006487 NM_005984 NM_001085427 NM_013236 NM_015140
NM_138415 NM_001195072 NM_001164501 NM_014339 NM_152868 NM_033257
NM_014941 NM_033386 NM_007311 NM_017911 NM_007098 NM_001670
NM_015653 NM_032775 NM_001172688 NM_001136029 NM_001024939 NM_002969
NM_052906 NM_152511 NM_001001479
```

Cluster 5, exemplar NM\_052945:

```
NM_001102371 NM_017829 NM_020243 NM_002305 NM_030882 NM_000496
NM_022720 NM_024053 NM_052945 NM_018006 NM_014433 NM_032561
NM_001142964 NM_181492 NM_003073 NM_015366 NM_005877 NM_148674
NM_005008 NM_017414 NM_000185 NM_001135911 NM_001199562 NM_003935
NM_003560 NM_001166242 NM_001165877 NM_004900 NM_001097 NM_174975
NM_004377 NM_001099294 NM_004121 NM_001146288 NM_002415 NM_001159546
NM_004327 NM_152426 NM_004861 NM_001193414 NM_001145398 NM_015715
NM_021974 NM_001159554 NM_001171660 NM_015124 NM_006932 NM_152906
NM_001002034 NM_000754 NM_145343
```

The final heatmap looks as follows:





a lot more different similarity measures. The user can make use of all variants implemented in `dist()` by using the `options` method (selects a distance measure) and `p` (specifies the exponent for the Minkowski distance, otherwise it is void) that are passed on to `dist()`. Presently, `dist()` provides the following variants of computing the distance  $d(\mathbf{x}, \mathbf{y})$  of two data samples  $\mathbf{x} = (x_1, \dots, x_n)$  and  $\mathbf{y} = (y_1, \dots, y_n)$ :

**Euclidean:**

$$d(\mathbf{x}, \mathbf{y}) = \sqrt{\sum_{i=1}^n (x_i - y_i)^2}$$

use `method="euclidean"` or do not specify argument `method` (since this is the default);

**Maximum:**

$$d(\mathbf{x}, \mathbf{y}) = \max_{i=1}^n |x_i - y_i|$$

use `method="maximum"`;

**Sum of absolute distances / Manhattan:**

$$d(\mathbf{x}, \mathbf{y}) = \sum_{i=1}^n |x_i - y_i|$$

use `method="manhattan"`;

**Canberra:**

$$d(\mathbf{x}, \mathbf{y}) = \sum_{i=1}^n \frac{|x_i - y_i|}{|x_i + y_i|}$$

summands with zero denominators are not taken into account; use `method="canberra"`;

**Minkowski:**

$$d(\mathbf{x}, \mathbf{y}) = \left( \sum_{i=1}^n (x_i - y_i)^p \right)^{\frac{1}{p}}$$

use `method="minkowski"` and specify  $p$  using the additional argument `p` (default is  $p=2$ , resulting in the standard Euclidean distance);

We do not consider `method="binary"` here, since it is irrelevant for real-valued data.

The function `negDistMat()` takes the distances computed with one of the variants listed above and returns  $-1$  times the  $r$ -th power of it, i.e.,

$$s(\mathbf{x}, \mathbf{y}) = -d(\mathbf{x}, \mathbf{y})^r. \quad (1)$$

The exponent  $r$  can be adjusted with the argument `r`. The default is  $r=1$ , hence, one has to supply  $r=2$  as in the above example to obtain squared distances.

Here are some examples. We use the corners of the two-dimensional unit square and its middle point  $(\frac{1}{2}, \frac{1}{2})$  as sample data:

```
> ex <- matrix(c(0,0,1,0,0.5,0.5,0,1,1,1),5,2,byrow=TRUE)
> ex
```

```

      [,1] [,2]
[1,]  0.0  0.0
[2,]  1.0  0.0
[3,]  0.5  0.5
[4,]  0.0  1.0
[5,]  1.0  1.0

```

Standard Euclidean distance:

```
> negDistMat(ex)
```

```

      1      2      3      4      5
1  0.0000000 -1.0000000 -0.7071068 -1.0000000 -1.4142136
2 -1.0000000  0.0000000 -0.7071068 -1.4142136 -1.0000000
3 -0.7071068 -0.7071068  0.0000000 -0.7071068 -0.7071068
4 -1.0000000 -1.4142136 -0.7071068  0.0000000 -1.0000000
5 -1.4142136 -1.0000000 -0.7071068 -1.0000000  0.0000000

```

Squared Euclidean distance:

```
> negDistMat(ex, r=2)
```

```

      1      2      3      4      5
1  0.0 -1.0 -0.5 -1.0 -2.0
2 -1.0  0.0 -0.5 -2.0 -1.0
3 -0.5 -0.5  0.0 -0.5 -0.5
4 -1.0 -2.0 -0.5  0.0 -1.0
5 -2.0 -1.0 -0.5 -1.0  0.0

```

Maximum norm-based distance:

```
> negDistMat(ex, method="maximum")
```

```

      1      2      3      4      5
1  0.0 -1.0 -0.5 -1.0 -1.0
2 -1.0  0.0 -0.5 -1.0 -1.0
3 -0.5 -0.5  0.0 -0.5 -0.5
4 -1.0 -1.0 -0.5  0.0 -1.0
5 -1.0 -1.0 -0.5 -1.0  0.0

```

Sum of absolute distances (aka Manhattan distance):

```
> negDistMat(ex, method="manhattan")
```

```

      1  2  3  4  5
1  0 -1 -1 -1 -2
2 -1  0 -1 -2 -1
3 -1 -1  0 -1 -1
4 -1 -2 -1  0 -1
5 -2 -1 -1 -1  0

```

Canberra distance:

```
> negDistMat(ex, method="canberra")
```

```

      1      2      3      4      5
1  0 -2.000000 -2.000000 -2.000000 -2.000000
2 -2  0.000000 -1.333333 -2.000000 -1.000000
3 -2 -1.333333  0.000000 -1.333333 -0.666667
4 -2 -2.000000 -1.333333  0.000000 -1.000000
5 -2 -1.000000 -0.666667 -1.000000  0.000000

```

Minkowski distance for  $p = 3$  (3-norm):

```
> negDistMat(ex, method="minkowski", p=3)
```

```

      1      2      3      4      5
1  0.0000000 -1.0000000 -0.6299605 -1.0000000 -1.2599210
2 -1.0000000  0.0000000 -0.6299605 -1.2599210 -1.0000000
3 -0.6299605 -0.6299605  0.0000000 -0.6299605 -0.6299605
4 -1.0000000 -1.2599210 -0.6299605  0.0000000 -1.0000000
5 -1.2599210 -1.0000000 -0.6299605 -1.0000000  0.0000000

```

## 7.2 Other similarity measures

The package `apcluster` offers three more functions for creating similarity matrices for real-valued data:

**Exponential transformation of distances:** the function `expSimMat()` is another wrapper around the standard function `dist()`. The difference is that, instead of the transformation (1), it uses the following transformation:

$$s(\mathbf{x}, \mathbf{y}) = \exp \left( - \left( \frac{d(\mathbf{x}, \mathbf{y})}{w} \right)^r \right)$$

Here the default is  $r=2$ . It is clear that  $r=2$  in conjunction with `method="euclidean"` results in the well-known *Gaussian kernel* / *RBF kernel* [4, 9, 11], whereas  $r=1$  in conjunction with `method="euclidean"` results in the similarity measure that is sometimes called *Laplace kernel* [4, 9]. Both variants (for non-Euclidean distances as well) can also be interpreted as *fuzzy equality/similarity relations* [2].

**Linear scaling of distances with truncation:** the function `linSimMat()` uses the transformation

$$s(\mathbf{x}, \mathbf{y}) = \max \left( 1 - \frac{d(\mathbf{x}, \mathbf{y})}{w}, 0 \right)$$

which is also often interpreted as a *fuzzy equality/similarity relation* [2].

**Linear kernel:** scalar products can also be interpreted as similarity measures, a view that is often adopted by kernel methods in machine learning. In order to provide the user with this option as well, the function `linKernel()` is available. For two data samples  $\mathbf{x} = (x_1, \dots, x_n)$  and  $\mathbf{y} = (y_1, \dots, y_n)$ , it computes the similarity as

$$s(\mathbf{x}, \mathbf{y}) = \sum_{i=1}^n x_i \cdot y_i.$$

The function has one additional argument, `normalize` (by default `FALSE`). If `normalize=TRUE`, values are normalized to the range  $[-1, +1]$  in the following way:

$$s(\mathbf{x}, \mathbf{y}) = \frac{\sum_{i=1}^n x_i \cdot y_i}{\sqrt{(\sum_{i=1}^n x_i^2) \cdot (\sum_{i=1}^n y_i^2)}}$$

Entries for which at least one of the two factors in the denominator is zero are set to zero (however, the user should be aware that this should be avoided anyway).

For the same example data as above, we obtain the following for the RBF kernel:

```
> expSimMat(ex)
```

	1	2	3	4	5
1	1.0000000	0.3678794	0.6065307	0.3678794	0.1353353
2	0.3678794	1.0000000	0.6065307	0.1353353	0.3678794
3	0.6065307	0.6065307	1.0000000	0.6065307	0.6065307
4	0.3678794	0.1353353	0.6065307	1.0000000	0.3678794
5	0.1353353	0.3678794	0.6065307	0.3678794	1.0000000

Laplace kernel:

```
> expSimMat(ex, r=1)
```

	1	2	3	4	5
1	1.0000000	0.3678794	0.4930687	0.3678794	0.2431167
2	0.3678794	1.0000000	0.4930687	0.2431167	0.3678794
3	0.4930687	0.4930687	1.0000000	0.4930687	0.4930687
4	0.3678794	0.2431167	0.4930687	1.0000000	0.3678794
5	0.2431167	0.3678794	0.4930687	0.3678794	1.0000000

Linear scaling of distances with truncation:

```
> linSimMat(ex, w=1.2)
```

```

      1      2      3      4      5
1 1.0000000 0.1666667 0.4107443 0.1666667 0.0000000
2 0.1666667 1.0000000 0.4107443 0.0000000 0.1666667
3 0.4107443 0.4107443 1.0000000 0.4107443 0.4107443
4 0.1666667 0.0000000 0.4107443 1.0000000 0.1666667
5 0.0000000 0.1666667 0.4107443 0.1666667 1.0000000
```

Linear kernel (we exclude (0,0)):

```
> linKernel(ex[2:5,])
```

```

      [,1] [,2] [,3] [,4]
[1,]  1.0  0.5  0.0   1
[2,]  0.5  0.5  0.5   1
[3,]  0.0  0.5  1.0   1
[4,]  1.0  1.0  1.0   2
```

Normalized linear kernel (we exclude (0,0)):

```
> linKernel(ex[2:5,], normalize=TRUE)
```

```

      [,1]      [,2]      [,3]      [,4]
[1,] 1.0000000 0.7071068 0.0000000 0.7071068
[2,] 0.7071068 1.0000000 0.7071068 1.0000000
[3,] 0.0000000 0.7071068 1.0000000 0.7071068
[4,] 0.7071068 1.0000000 0.7071068 1.0000000
```

## 8 Miscellaneous

### 8.1 Clustering named objects

The function `apcluster()` and all functions for computing distance matrices are implemented to recognize names of data objects and to correctly pass them through computations. The mechanism is best described with a simple example:

```

> x3 <- c(1, 2, 3, 7, 8, 9)
> names(x3) <- c("a", "b", "c", "d", "e", "f")
> s3 <- negDistMat(x3, r=2)
```

So we see that the `names` attribute must be used if a vector of named one-dimensional samples is to be clustered. If the data are not one-dimensional (a matrix instead), object names must be stored in the row names of the data matrix.

All functions for computing similarity matrices recognize the object names. The resulting similarity matrix has the list of names both as row and column names.

```
> s3
```

```
      a   b   c   d   e   f
a    0  -1  -4 -36 -49 -64
b   -1   0  -1 -25 -36 -49
c   -4  -1   0 -16 -25 -36
d  -36 -25 -16   0  -1  -4
e  -49 -36 -25  -1   0  -1
f  -64 -49 -36  -4  -1   0
```

```
> colnames(s3)
```

```
[1] "a" "b" "c" "d" "e" "f"
```

The function `apcluster()` and all related functions use column names of similarity matrices as object names. If object names are available, clustering results are by default shown by names.

```
> apres3a <-apcluster(s3)
```

```
> apres3a
```

APResult object

```
Number of samples      = 6
Number of iterations   = 124
Input preference       = -25
Sum of similarities    = -4
Sum of preferences    = -50
Net similarity         = -54
Number of clusters     = 2
```

Exemplars:

```
  b e
```

Clusters:

```
  Cluster 1, exemplar b:
```

```
    a b c
```

```
  Cluster 2, exemplar e:
```

```
    d e f
```

```
> apres3a@exemplars
```

```
b e
```

```
2 5
```

```
> apres3a@clusters
```

```
[[1]]  
a b c  
1 2 3
```

```
[[2]]  
d e f  
4 5 6
```

## 8.2 Computing a label vector from a clustering result

For later classification or comparisons with other clustering methods, it may be useful to compute a label vector from a clustering result. Our package provides an instance of the generic function `labels()` for this task. As obvious from the following example, the argument `type` can be used to determine how to compute the label vector.

```
> apres3a@exemplars  
  
b e  
2 5  
  
> labels(apres3a, type="names")  
  
[1] "b" "b" "b" "e" "e" "e"  
  
> labels(apres3a, type="exemplars")  
  
[1] 2 2 2 5 5 5  
  
> labels(apres3a, type="enum")  
  
[1] 1 1 1 2 2 2
```

The first choice, "names" (default), uses names of exemplars as labels (if names are available, otherwise an error message is displayed). The second choice, "exemplars", uses indices of exemplars (enumerated as in the original data set). The third choice, "enum", uses indices of clusters (consecutively numbered as stored in the slot `clusters`; analogous to the standard implementation of `cutree()` or the `clusters` field of the list returned by the standard function `kmeans()`).

## 8.3 Implementation and performance issues

Prior to version 1.2.0, `apcluster()` was implemented in R. Starting with version 1.2.0, the main iteration loop of `apcluster()` has been implemented in C++ using the Rcpp package [3], which has led to a speedup in the range of a factor of 9–10.

Note that `details=TRUE` requires quite an amount of additional memory. If possible, avoid this for larger data sets.

The asymptotic computational complexity of `aggExCluster()` is  $\mathcal{O}(l^3)$  (where  $l$  is the number of samples or clusters from which the clustering starts). This may result in excessively long computation times if `aggExCluster()` is used for larger data sets without using affinity propagation first. For real-world data sets, in particular, if they are large, we recommend to use affinity propagation first and then, if necessary, to use `aggExCluster()` to create a cluster hierarchy.

## 9 Future Extensions

We currently have no implementation that exploits sparsity of similarity matrices. The implementation of *sparse AP* and *leveraged AP* which are available as Matlab code from the AP Web page<sup>1</sup> is left for future extensions of the package. Presently, we only offer a function `sparseToFull()` that converts similarity matrices from sparse format into a full  $l \times l$  matrix.

## 10 Change Log

### Version 1.2.0:

- reimplementation of `apcluster()` in C++ using the Rcpp package [3] which reduces computation times by a factor of 9-10
- obsolete function `apclusterLM()` removed
- updates of help pages and vignette

### Version 1.1.1:

- updated citation
- minor corrections in help pages and vignette

### Version 1.1.0:

- added exemplar-based agglomerative clustering function `aggExCluster()`
- added various plotting functions for dendrograms and heatmaps
- extended help pages and vignette according to new functionality
- added sequence analysis example to vignette along with data set `ch22Promoters`
- re-organization of variable names in vignette
- added option `verbose` to `apclusterK()`
- numerous minor corrections in help pages and vignette

### Version 1.0.3:

- Makefile in `inst/doc` eliminated to avoid installation problems
- renamed vignette to “`apcluster`”



**Version 1.0.2:**

- replacement of computation of responsibilities and availabilities in function `apcluster()` by pure matrix operations (see 8.3 above); traditional implementation à la Frey and Dueck still available as function `apclusterLM`;
- improved support for named objects (see 8.1)
- new function for computing label vectors (see 8.2)
- re-organization of package source files and help pages

**Version 1.0.1:** first official release, released March 2, 2010

## 11 How to Cite This Package

If you use this package for research that is published later, you are kindly asked to cite it as follows:

U. Bodenhofer, A. Kothmeier, and S. Hochreiter (2011). APCluster: an R package for affinity propagation clustering. *Bioinformatics* **27**(17):2463–2464. DOI: 10.1093/bioinformatics/btr406.

Moreover, we insist that, any time you cite the package, you also cite the original paper in which affinity propagation has been introduced [5].

To obtain BibTeX entries of the two references, you can enter the following into your R session:

```
> toBibtex(citation("apcluster"))
```

## References

- [1] U. Bodenhofer, A. Kothmeier, and S. Hochreiter. APCluster: an R package for affinity propagation clustering. *Bioinformatics*, 27(17):2463–2464, 2011.
- [2] B. De Baets and R. Mesiar. Metrics and  $T$ -equalities. *J. Math. Anal. Appl.*, 267:531–547, 2002.
- [3] D. Eddelbuettel and R. François. Rcpp: seamless R and C++ integration. *J. Stat. Softw.*, 40(8):1–18, 2011.
- [4] C. H. FitzGerald, C. A. Micchelli, and A. Pinkus. Functions that preserve families of positive semidefinite matrices. *Linear Alg. Appl.*, 221:83–102, 1995.
- [5] B. J. Frey and D. Dueck. Clustering by passing messages between data points. *Science*, 315(5814):972–976, 2007.
- [6] A. K. Jain, M. N. Murty, and P. J. Flynn. Data clustering: a review. *ACM Comput. Surv.*, 31(3):264–323, 1999.

- [7] A. Karatzoglou, A. Smola, K. Hornik, and A. Zeileis. kernlab – an S4 package for kernel methods in R. *J. Stat. Softw.*, 11(9):1–20, 2004.
- [8] C. Leslie, E. Eskin, and W. S. Noble. The spectrum kernel: a string kernel for SVM protein classification. In R. B. Altman, A. K. Dunker, L. Hunter, K. Lauderdale, and T. E. D. Klein, editors, *Pacific Symposium on Biocomputing 2002*, pages 566–575. World Scientific, 2002.
- [9] C. A. Micchelli. Interpolation of scattered data: Distance matrices and conditionally positive definite functions. *Constr. Approx.*, 2:11–22, 1986.
- [10] R. S. Michalski and R. E. Stepp. Clustering. In S. C. Shapiro, editor, *Encyclopedia of artificial intelligence*, pages 168–176. John Wiley & Sons, Chichester, 1992.
- [11] B. Schölkopf and A. J. Smola. *Learning with Kernels*. Adaptive Computation and Machine Learning. MIT Press, Cambridge, MA, 2002.
- [12] J. H. Ward Jr. Hierarchical grouping to optimize an objective function. *J. Amer. Statist. Assoc.*, 58:236–244, 1963.