

Graphical Independence Networks

A vignette for the gRain package

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1 Bayesian networks

1.1 Introduction

The gRain package implements Bayesian Networks (hereafter often abbreviated BNs). The name gRain is an acronym for [gra]phical [i]ndependence [n]etworks. The main reference for gRain is Højsgaard (2012), see also `citation("gRain")`.

Moreover, Højsgaard, Edwards, and Lauritzen (2012) gives a broad treatment of graphical models (including Bayesian networks) More information about the package, other graphical modelling packages and development versions is available from

(<http://people.math.aau.dk/~sorenh/software/gR>)

1.1.1 Example: Chest clinic

This section reviews the chest clinic example of Lauritzen and Spiegelhalter (1988) (illustrated in Figure @ref(fig:chest-LS)) and shows one way of specifying the model in gRain. Lauritzen and Spiegelhalter (1988) motivate the chest clinic example with the following narrative:

“Shortness-of-breath (dyspnoea) may be due to tuberculosis, lung cancer or bronchitis, or none of them, or more than one of them. A recent visit to Asia increases the chances of tuberculosis, while smoking is known to be a risk factor for both lung cancer and bronchitis. The results of a single chest X-ray do not discriminate between lung cancer and tuberculosis, as neither does the presence or absence of dyspnoea.”

```
chest_dag <- dag(list("asia", c("tub", "asia"), "smoke", c("lung", "smoke"), c("bronc",
"smoke"), c("either", "tub", "lung"), c("xray", "either"), c("dysp",
"bronc", "either")))
library(igraph)
par(mar=c(0,0,0,0))
plot(chest_dag)
```

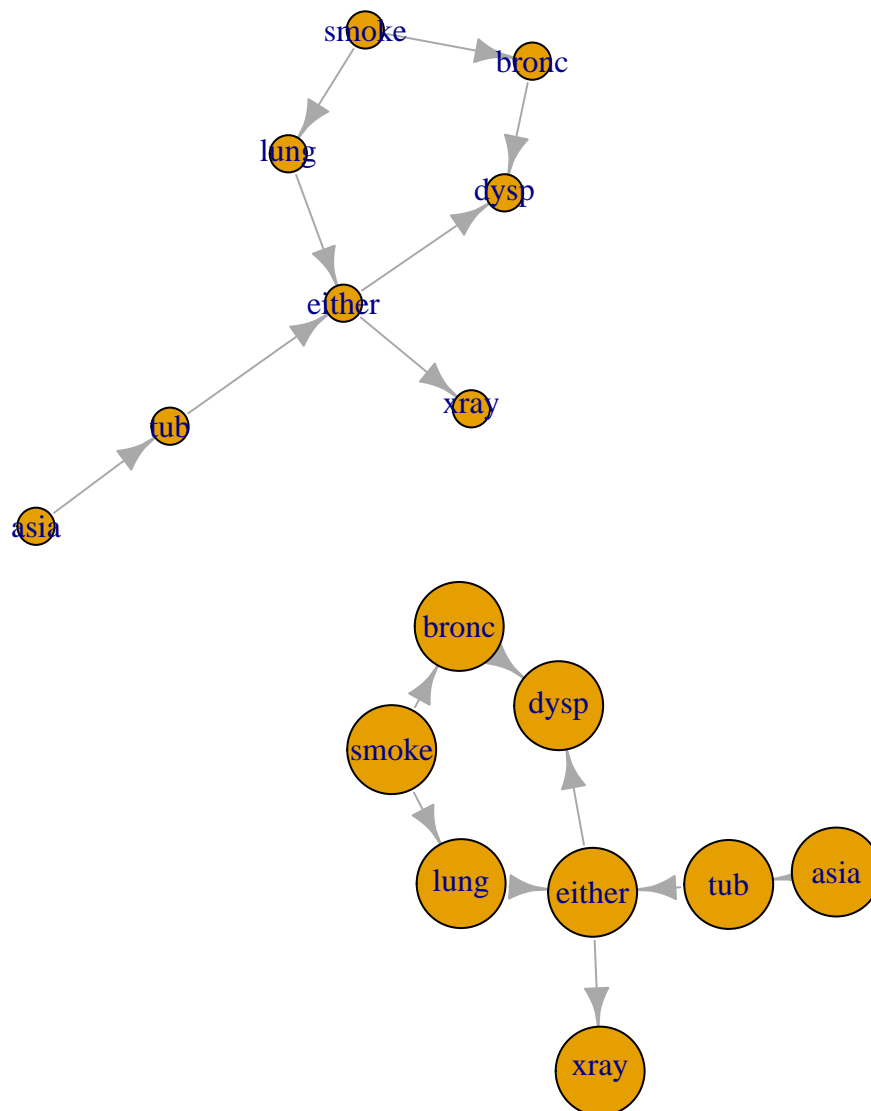


Figure 1: Chest clinic example from Lauritzen and Spiegelhalter (1988).

1.2 Building a Bayesian network

The description above involves the following binary variables: a = asia, s = smoker, t = tuberculosis, l = lung cancer, b = bronchitis, e = either tuberculosis or lung cancer, d = dyspnoea and x = xray.

Each variable is binary and can take the values “yes” and “no”: Note that e is a logical variable which is true (yes) if either t or l are true (yes) and false (no) otherwise. The connection between the variables is displayed by the DAG (directed acyclic graph) in Figure~@ref(fig:chest-LS).

A joint probability density factorizing according to a DAG with nodes V can be constructed as follows:

Each node $v \in V$ has a set $pa(v)$ of parents and each node $v \in V$ has a finite set of states. A joint distribution over the variables V can be given as a product of conditional distributions

$$(\#eq : dagfact1)p(V) = \prod_{v \in V} p(v|pa(v)) \quad (1)$$

where $p(v|pa(v))$ is a function defined on $(v, pa(v))$. This function satisfies that

$$\sum_{v^*} p(v = v^*|pa(v)) = 1,$$

i.e. that for each configuration of the parents $pa(v)$, the sum over the levels of v equals one. Hence $p(v|pa(v))$ becomes the conditional distribution of v given $pa(v)$. In practice $p(v|pa(v))$ is specified as a table called a conditional probability table or a CPT for short. Thus, a Bayesian network can be regarded as a complex stochastic model built up by putting together simple components (conditional probability distributions). A joint probability density for all eight variables in Figure~@ref(fig:chest-LS) can be constructed as

$$(\#eq : chestfact1)p(V) = p(a)p(s)p(t|a)p(l|s)p(b|s)p(e|t, l)p(d|e, b)p(x|e). \quad (2)$$

1.3 Queries to networks

Suppose we are given the evidence (sometimes also called “finding”) that a set of variables $E \subset V$ have a specific value e^* . With this evidence, we are often interested in the conditional distribution $p(v|E = e^*)$ for some of the variables $v \in V \setminus E$ or in $p(U|E = e^*)$ for a set $U \subset V \setminus E$. Interest might also be in calculating the probability of a specific event, e.g. the probability of seeing a specific evidence, i.e. $p(E = e^*)$. Other types of evidence (called soft evidence, virtual evidence or likelihood evidence) are discussed in Section @ref(sec:hard-soft).

For example that a person has recently visited Asia and suffers from dyspnoea, i.e. $a = \text{yes}$ and $d = \text{yes}$. In the chest clinic example, interest might be in $p(l|e^*)$, $p(t|e^*)$ and $p(b|e^*)$, or possibly in the joint (conditional) distribution $p(l, t, b|e^*)$.

1.4 A one-minute version of gRain

1.4.1 Specifying a Bayesian network

A simple way of specifying the model for the chest clinic example is as follows.

Specify conditional probability tables (with values as given in Lauritzen and Spiegelhalter (1988)) (there are other ways of specifying conditional probability tables, see the package documentation):

```
yn <- c("yes", "no")
a <- cpt(~asia, values=c(1, 99), levels=yn)
t.a <- cpt(~tub|asia, values=c(5, 95, 1, 99), levels=yn)
s <- cpt(~smoke, values=c(5, 5), levels=yn)
l.s <- cpt(~lung|smoke, values=c(1, 9, 1, 99), levels=yn)
b.s <- cpt(~bronc|smoke, values=c(6, 4, 3, 7), levels=yn)
e.lt <- cpt(~either|lung:tub, values=c(1, 0, 1, 0, 1, 0, 0, 1), levels=yn)
x.e <- cpt(~xray|either, values=c(98, 2, 5, 95), levels=yn)
d.be <- cpt(~dysp|bronc:either, values=c(9, 1, 7, 3, 8, 2, 1, 9), levels=yn)
```

Compile list of conditional probability tables.

```
chest_cpt <- compile_cpt(a, t.a, s, l.s, b.s, e.lt, x.e, d.be)
chest_cpt
```

```
## P( asia )
## P( tub | asia )
## P( smoke )
## P( lung | smoke )
## P( bronc | smoke )
## P( either | lung tub )
## P( xray | either )
## P( dysp | bronc either )
```

The components are arrays, but coercion into dataframes sometimes makes it easier to digest the components.

```
chest_cpt$tub
```

```
##      asia
## tub   yes   no
##   yes 0.05 0.01
##   no  0.95 0.99
```

```
chest_cpt$tub |> as.data.frame.table()
```

```
##   tub asia Freq
## 1 yes  yes  0.05
## 2 no   yes  0.95
## 3 yes  no   0.01
## 4 no   no   0.99
```

Create the network:

```
chest_bn <- grain(chest_cpt)
chest_bn
```

```
## Independence network: Compiled: TRUE Propagated: FALSE Evidence: FALSE
```

Default is that the network is compiled at creation time, but if one chooses not to do so, compilation can be done with:

```
chest_bn <- compile(chest_bn)
```

1.4.2 Setting evidence and querying a network

In the chest clinic example, there are three disease variables, two background variables and two symptoms. Following the narrative, we can set evidence that a person has recently visited Asia and has dyspnoea:

```
disease <- c("tub", "lung", "bronc")
asia_dysp <- list(asia="yes", dysp="yes")
```

Initially, the network can be queried without evidence:

```
chest_bn |> qgrain(nodes=disease, type="marginal")
```

```
## $tub
## tub
##   yes   no
## 0.0104 0.9896
##
## $lung
## lung
##   yes   no
## 0.055 0.945
##
## $bronc
## bronc
##   yes   no
```

```
## 0.45 0.55
chest_bn |> qgrain(nodes=disease, type="marginal", simplify = TRUE)

##           yes      no
## tub    0.0104 0.9896
## lung   0.0550 0.9450
## bronc  0.4500 0.5500

chest_bn |> qgrain(nodes=disease, type="joint")

## , , bronc = yes
##
##      lung
## tub      yes      no
## yes 0.0003276 0.004352
## no  0.0311724 0.414148
##
## , , bronc = no
##
##      lung
## tub      yes      no
## yes 0.0002444 0.005476
## no  0.0232556 0.521024

chest_bn |> qgrain(nodes=disease, type="joint", simplify = TRUE)

##  tub lung bronc      Freq
## 1 yes  yes   yes 0.0003276
## 2 no  yes   yes 0.0311724
## 3 yes  no    yes 0.0043524
## 4 no  no    yes 0.4141476
## 5 yes  yes   no  0.0002444
## 6 no  yes   no  0.0232556
## 7 yes  no    no  0.0054756
## 8 no  no    no  0.5210244

chest_bn |> qgrain(nodes=disease, type="conditional")

## , , bronc = yes
##
##      lung
## tub      yes      no
## yes 0.0104 0.0104
## no  0.9896 0.9896
##
## , , bronc = no
##
##      lung
## tub      yes      no
## yes 0.0104 0.0104
## no  0.9896 0.9896

chest_bn |> qgrain(nodes=disease, type="conditional", simplify = TRUE)

##  tub lung bronc      Freq
## 1 yes  yes   yes 0.0104
## 2 no  yes   yes 0.9896
## 3 yes  no    yes 0.0104
## 4 no  no    yes 0.9896
## 5 yes  yes   no  0.0104
## 6 no  yes   no  0.9896
```

```
## 7 yes    no    no 0.0104
## 8 no     no    no 0.9896
```

Above we obtain the marginal, joint distributions and conditional distribution for the disease variables. The output can in some cases be simplified to dataframes. For the conditional distribution, we obtain the conditional distribution of the first node given the rest of the nodes.

1.5 Setting evidence

Evidence can be entered in different ways:

```
asia_dysp <- list(asia="yes", dysp="yes")
chest_ev <- chest_bn |>
  setEvidence(evidence=asia_dysp)
## chest_ev <- chest_bn |> ## For backward compatibility
##   setEvidence(nodes=c("asia", "dysp"), states=c("yes", "yes"))
```

The evidence is a list and can conveniently be displayed as a dataframe:

```
chest_ev |> getEvidence() |> as.data.frame()

##   nodes is_hard hard_state evi_weight
## 1  asia    TRUE         yes         1, 0
## 2  dysp    TRUE         yes         1, 0

chest_ev |> querygrain(nodes=disease, simplify = TRUE)
```

```
##           yes    no
## tub   0.08775 0.9122
## lung  0.09953 0.9005
## bronc 0.81140 0.1886
```

```
chest_ev |> pEvidence()
```

```
## [1] 0.004501
```

The network can be queried again, and we can also obtain the probability of observing this evidence:

```
chest_ev |> querygrain(nodes=disease, simplify=TRUE)
```

```
##           yes    no
## tub   0.08775 0.9122
## lung  0.09953 0.9005
## bronc 0.81140 0.1886
```

```
chest_ev |> pEvidence()
```

```
## [1] 0.004501
```

```
chest_bn |> pEvidence(evidence=list(asia="yes", dysp="yes"))
```

```
## [1] 0.004501
```

The probability of observing a specific evidence can be found by setting the evidence as a vector of weights. This is useful in connection with soft evidence (also called likelihood evidence), see Section~@ref(sec:hard-soft).

1.6 Hints and shortcuts

An alternative way of specifying a network is by first defining CPTs and then entering values afterwards. Programmatically, this can be done as:

```
yn <- c("yes", "no")

node_parents_list <-
```

```

list("asia", c("tub", "asia"), "smoke", c("lung", "smoke"),
     c("bronc", "smoke"), c("either", "tub", "lung"),
     c("xray", "either"), c("dysp", "bronc", "either"))

## nms <- unique(unlist(node_parents_list))
## universe <- setNames(rep(list(yn),length(nms)), nms)
## universe /> head(3)

chest_dummy_cpt2 <- lapply(node_parents_list, function(f){
  cpt(f, levels=yn)
})
bn_template <- compile_cpt(chest_dummy_cpt2) |> grain()

```

Above the network has ones in all potentials. Next update values in (some of the) potentials:

```

cpt_values <- list(asia=c(1, 99),
                  tub=c(5, 95, 1, 99),
                  smoke=c(5, 5),
                  lung=c(1, 9, 1, 99),
                  bronc=c(6, 4, 3, 7),
                  either=c(1, 0, 1, 0, 1, 0, 0, 1),
                  xray=c(98, 2, 5, 95),
                  dysp=c(9, 1, 7, 3, 8, 2, 1, 9)
                  )

bn_real <- replace_cpt(bn_template, cpt_values)

bn_template |> qgrain(evi=asia_dysp, nodes=disease, simplify = TRUE)

##          yes  no
## tub    0.5 0.5
## lung   0.5 0.5
## bronc  0.5 0.5

bn_real |> qgrain(evi=asia_dysp, nodes=disease, simplify = TRUE)

```

```

##          yes    no
## tub    0.08775 0.9122
## lung   0.09953 0.9005
## bronc  0.81140 0.1886

```

Consider querying a network where focus is on marginal distributions (the default). If all variables have the same levels (as the case is here), the output can be coerced to a dataframe:

```

querygrain(chest_bn, nodes=disease, simplify = TRUE)

##          yes    no
## tub    0.0104 0.9896
## lung   0.0550 0.9450
## bronc  0.4500 0.5500

```

In the more general case the output can be coerced to a list of dataframes

```

querygrain(chest_bn, nodes=disease, result="data.frame")

## $tub
## tub
##    yes    no
## 0.0104 0.9896
##
## $lung
## lung

```

```
##      yes      no
## 0.055 0.945
##
## $bronc
## bronc
##      yes      no
## 0.45 0.55
```

A typical use of gRain involves setting evidence and then querying the network afterwards. This can all be done in one call of `querygrain()` (notice that this does not alter the network object):

```
chest_bn |> querygrain(evidence=asia_dysp,
                      nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.08775 0.9122
## lung   0.09953 0.9005
## bronc  0.81140 0.1886
```

Evidence can also be given as a vector of weights.

```
chest_bn |> querygrain(evidence=list(asia=c(1, 0), dysp=c(1, 0)),
                      nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.08775 0.9122
## lung   0.09953 0.9005
## bronc  0.81140 0.1886
```

The weights must be non-negative but need not sum to one. This is important in connection with soft evidence (also called likelihood evidence), see Section~@ref(sec:hard-soft). Above, the weights could also have been set as `c(1, 0)`. The important part is that the zero excludes certain states as being impossible.

Nodes on which evidence is given are not reported unless `exclude=FALSE`

```
querygrain(chest_bn,
            evidence=list(asia=c(1, 0), dysp=c(1, 0)),
            nodes=c("lung", "bronc", "asia", "dysp"),
            exclude=FALSE, simplify = TRUE)
```

```
##           yes      no
## lung   0.09953 0.9005
## bronc  0.81140 0.1886
## asia   1.00000 0.0000
## dysp   1.00000 0.0000
```

If `nodes` are not specified, queries for all nodes without evidence are reported.

```
querygrain(chest_bn,
            evidence=asia_dysp,
            simplify = TRUE)
```

```
##           yes      no
## NA           NA      NA
## tub    0.08775 0.9122
## smoke  0.62592 0.3741
## lung   0.09953 0.9005
## bronc  0.81140 0.1886
## either 0.18230 0.8177
## xray   0.21954 0.7805
## NA.1           NA      NA
```

If `nodes` are not specified and `exclude=FALSE`, then queries for all nodes are reported.


```
querygrain(chest_bn,
           evidence=asia_dysp,
           exclude = FALSE, simplify = TRUE)
```

```
##           yes      no
## asia    1.00000 0.0000
## tub     0.08775 0.9122
## smoke   0.62592 0.3741
## lung    0.09953 0.9005
## bronc   0.81140 0.1886
## either  0.18230 0.8177
## xray    0.21954 0.7805
## dysp    1.00000 0.0000
```

1.7 Conditioning on evidence with zero probability

Consider setting the evidence

```
chest_bn3 <- setEvidence(chest_bn, evidence=list(either="no", tub="yes"))
```

Under the model, this specific evidence has zero probability: `either` is true if `tub` is true or `lung` is true (or both). Hence the specific evidence is impossible and therefore, all conditional probabilities are (under the model) undefined:

```
pEvidence(chest_bn3)
```

```
## [1] 0
```

```
querygrain(chest_bn3, nodes=disease, type="joint")
```

```
##      bronc
## lung yes no
## yes NaN NaN
## no  NaN NaN
```

1.8 Hard and soft (likelihood) evidence

Below we describe how to work with virtual evidence (also known as soft evidence or likelihood evidence) in gRain. This is done via the function `setEvidence()`.

The clique potential representation in a Bayesian network gives

$$p(x) \propto \psi(x) = \prod_C \psi_C(x_C). \quad (3)$$

Here we recall that the whole idea in computations with Bayesian networks is to avoid calculation the product on the right hand side above. Instead computations are based on propagation (multiplying, dividing and summing clique potentials ψ_C in an appropriate order, and such an appropriate order comes from a junction tree). The normalizing constant, say $c = \sum_x \psi(x)$, comes out of propagation as a “by-product”.

Suppose a set of nodes E are known to have a specific value, i.e. $x_E = x_E^*$. This is called hard evidence. The probability of the event $x_E = x_E^*$ is

$$p(x_E = x_E^*) = E_p\{I(x_E = x_E^*)\} = \sum_x I(x_E = x_E^*)p(x) = \frac{1}{c} \sum_x I(x_E = x_E^*)\psi(x) \quad (4)$$

The computations are based on modifying the clique potentials ψ_C by giving value zero to states in ψ_C which are not consistent with $x_E = x_E^*$. This can be achieved with an indicator function, say $L_C(x_C)$ such that we obtain a set of new potentials $\tilde{\psi}_C(x) = L_C(x_C)\psi_C(x_C)$. Propagation with these new potentials gives, as a by product, $\tilde{c} = \sum \tilde{\psi}(x)$ where $\psi(x) = \prod_C \psi_C(x_C)$. Consequently, we have $p(x_E = x_E^*) = \tilde{c}/c$.

In a more general setting we may have non-negative weights $L(x)$ for each value of x . We may calculate

$$E_p\{L(X)\} = \sum_x L(x)p(x) \quad (5)$$

If $L(X)$ factorizes as $L(X) = \prod_C L_C(X_C)$ then the computations are carried out as outlined above, i.e. by the message passing scheme.

1.8.1 Hard evidence

Suppose we want to make a diagnosis about tuberculosis given the evidence that a person is a smoker. We call such evidence hard evidence because the state of the variables are known with certainty. The function `setEvidence()` can be used for this purpose. The following forms are equivalent (the reason will be explained below):

```
chest_ev1 <- setEvidence(chest_bn, evidence=list(smoke="yes"))
chest_ev2 <- setEvidence(chest_bn, evidence=list(smoke=c(1, 0)))
```

```
chest_bn |> querygrain(nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.0550 0.9450
## bronc  0.4500 0.5500
```

```
chest_ev1 |> querygrain(nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.1000 0.9000
## bronc  0.6000 0.4000
```

1.8.2 Soft evidence (also called virtual evidence or likelihood evidence)

Suppose we are not certain if a patient is a smoker or not. We shall assume that for patients who are smokers, we would (correctly) guess so in 80% of the cases, whereas for patients who are not smokers we would (erroneously) guess that they are smokers in 10% of the cases.

In gRain this situation can be handled in two different ways. One way is to introduce a new variable `smoke_guess` with `smoke` as its only parent and then we enter evidence for this node.

```
g.s <- cpt(~ smoke_guess|smoke, levels=yn,
           values=c(.8, .2, .1, .9))
g.s
```

```
##           smoke
## smoke_guess yes  no
##           yes 0.8 0.1
##           no  0.2 0.9
```

```
chest_ext <- c(cpt_list, list(g.s)) |> compile_cpt() |> grain()
querygrain(chest_ext, nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.0550 0.9450
## bronc  0.4500 0.5500
```

```
querygrain(chest_ext, nodes=disease, evidence=list(smoke="yes"), simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.1000 0.9000
## bronc  0.6000 0.4000
```

```
querygrain(chest_ext, nodes=disease, evidence=list(smoke_guess="yes"), simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.0900 0.9100
## bronc  0.5667 0.4333
```

Another approach is to enter virtual evidence in the original network as shown below.

```
chest_ve <- chest_bn |>
  setEvidence(evidence = list(smoke = c(.8, .1)))
chest_ve |> querygrain(nodes=disease, simplify = TRUE)
```

```
##           yes      no
## tub    0.0104 0.9896
## lung   0.0900 0.9100
## bronc  0.5667 0.4333
```

```
getEvidence(chest_ve)
```

```
## $nodes
## [1] "smoke"
##
## $is_hard
## [1] FALSE
##
## $hard_state
## [1] NA
##
## $evi_weight
## $evi_weight[[1]]
## smoke
## yes no
## 0.8 0.1
##
##
## attr("class")
## [1] "grain_evidence" "list"
```

Consequently, specifying the hard evidence that `smoke="yes"` can be done as

```
chest_bn |> setEvidence(evidence=list(smoke=c(1, 0)))
```

```
## Independence network: Compiled: TRUE Propagated: TRUE Evidence: TRUE
```

1.9 Building a network from data

When building a network from data, there are two situations to be distinguished between. 1) The network is known and the data is used to estimate the parameters in the network. 2) The network is unknown and the data is used to estimate the network structure and the parameters in the network.

In both cases it is possible to have a network specified as a dag or as an undirected (chordal) graph. The following code illustrates how to build a network from data. The data is simulated data from the chest clinic example.

1.9.1 Building a network from a known network

The following list defines a dag for the chest clinic example: Each component is a list with two elements: the first element is the node and the second element is a vector of parents.

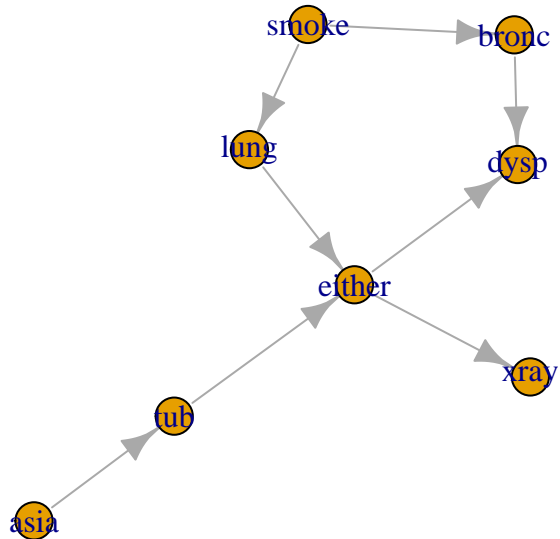
```
node_parents_list <- list("asia", c("tub", "asia"), "smoke", c("lung", "smoke"),
  c("bronc", "smoke"), c("either", "tub", "lung"),
```

```

c("xray", "either"), c("dysp", "bronc", "either"))

g1 <- dag(node_parents_list)
par(mar=c(0,0,0,0))
plot(g1)

```



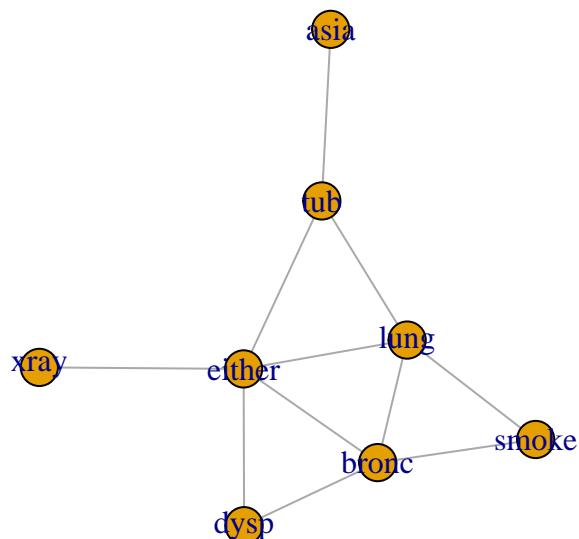
The following list defines an undirected graph for the chest clinic example: Each component is a list with the nodes in the clique.

```

cliq <- list(c("xray", "either"), c("asia", "tub"), c("smoke", "lung",
"bronc"), c("lung", "either", "tub"), c("lung", "either", "bronc"
), c("bronc", "either", "dysp"))

g2 <- ug(cliq)
par(mar=c(0,0,0,0))
plot(g2)

```



A network can be built from data using:

```

bn1 <- grain(g1, data=gRbase::chestSim100000)
bn2 <- grain(g2, data=gRbase::chestSim100000)

```

```

## NAs found in clique potential(s) for clique(s):
## c("bronc", "lung", "either")

```

```
## ... consider using the smooth argument
```

It is explained in the references that the two networks specify the same joint distribution because internally a dag is converted to an undirected chordal graph which provides the basis for the computations. In practice the two joint distributions differ slightly, and this is because the way in which information is extracted from data, see examples below.

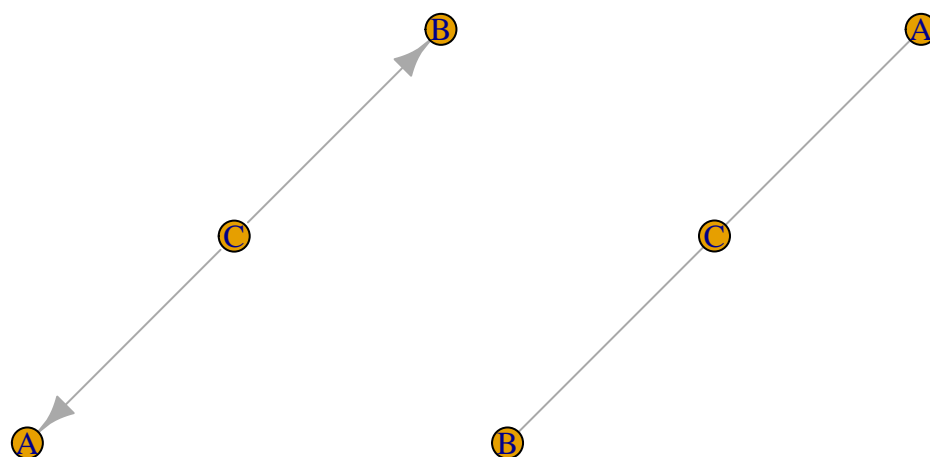
```
j1 <- querygrain(bn1, type="joint")
j2 <- querygrain(bn2, type="joint")
d <- j1 %a-% j2
max(abs(d))
```

```
## [1] NaN
```

1.10 Building networks from data

The following two graphs specify the same model stating that A and C are conditionally independent given B. The first graph is a directed acyclic graph (DAG) and the second graph is an undirected graph (UG).

```
g1 <- dag(~A:C + B:C, result="igraph")
g2 <- ug(~A:C + B:C, result="igraph")
par(mfrow=c(1,2), mar=c(0,0,0,0))
plot(g1); plot(g2)
```



Given such a graph, we can build a network from data. Suppose data is as follows:

```
tab <- tabNew(~A:B:C, levels=c("+", "-"),
              values=c(1, 2, 3, 5, 1, 2, 1, 4))
```

A network can be built from data using:

```
bn1 <- grain(g1, data=tab)
bn2 <- grain(g2, data=tab)
```

The two networks specify the same joint distribution:

```
j1 <- querygrain(bn1, type="joint")
j2 <- querygrain(bn2, type="joint")
d <- j1 %a-% j2
max(abs(d))
```

```
## [1] 2.776e-17
```

In the process of creating networks, conditional probability tables are extracted when the graph is a dag and clique potentials are extracted when the graph is a chordal (i.e. triangulated) undirected graph. This takes place as follows (internally):

Need to estimate $p(A|B)$, $p(C|B)$ and $p(B)$ from the data. This can be done as follows:

```
p <- extract_cpt(tab, g1) |> c()
p
```

```
## [[1]]
## C
##      +      -
## 0.5789 0.4211
##
## [[2]]
##      C
## A      +      -
## + 0.3636 0.25
## - 0.6364 0.75
##
## [[3]]
##      C
## B      +      -
## + 0.2727 0.375
## - 0.7273 0.625
```

Likewise, we can extract the clique potentials from the data, call these $q(AC)$ and $q(BC)$:

```
q <- extract_pot(tab, g2) |> c()
q
```

```
## [[1]]
##      C
## A      +      -
## + 0.2105 0.1053
## - 0.3684 0.3158
##
## [[2]]
##      C
## B      +      -
## + 0.2727 0.375
## - 0.7273 0.625
```

Clique potentials are not uniquely defined, but the product of clique potentials is. In the implementation in gRain, $q(AC)$ is the relative frequency in the AC -marginal table, i.e. $p(AB)$. Moreover $q(BC)$ is the frequency in the BC -marginal table divided by the frequency in the C -marginal table. Hence, the product of the clique potentials and the product of the cpts is the same as the joint distribution:

```
tabProd(p) |> ftable()
```

```
##      C      +      -
## A B
## + +  0.05742 0.03947
## -  0.15311 0.06579
## - +  0.10048 0.11842
## -  0.26794 0.19737
```

```
tabProd(q) |> ftable()
```

```
##      C      +      -
## A B
## + +  0.05742 0.03947
## -  0.15311 0.06579
## - +  0.10048 0.11842
## -  0.26794 0.19737
```

1.10.1 Handling zeros in the data

Next suppose data is as follows

```
tab0 <- tab
tab0[1:4] <- 0
tab0
```

```
## , , C = +
##
##      B
## A    + -
## + 0 0
## - 0 0
##
## , , C = -
##
##      B
## A    + -
## + 1 1
## - 2 4
```

In the process of creating networks above, the following quantities are computed:

```
n_BC <- tab0 |> tabMarg(~B:C)
n_AC <- tab0 |> tabMarg(~A:C)
n_C  <- tab0 |> tabMarg(~C)
```

```
p.B_C <- n_BC %a/% n_C
p.A_C <- n_AC %a/% n_C
p.C   <- n_C / sum(n_C)
p.C
```

```
## C
## + -
## 0 1
```

```
p.B_C
```

```
##      C
## B      +      -
## + NaN 0.375
## - NaN 0.625
```

```
p.A_C
```

```
##      C
## A      +      -
## + NaN 0.25
## - NaN 0.75
```

```
bn01 <- grain(g1, data=tab0)
```

```
## NAs found in cpt(s) for node(s): A, B
## ... consider using the smooth argument
```

```
bn02 <- grain(g2, data=tab0)
```

```
## NAs found in clique potential(s) for clique(s):
## c("B", "C")
## ... consider using the smooth argument
```

```
p <- extract_cpt(tab0, g1) |> c()
```

```
## NAs found in cpt(s) for node(s): A, B
```

```

## ... consider using the smooth argument
q <- extract_pot(tab0, g2) |> c()

## NAs found in clique potential(s) for clique(s):
## c("B", "C")
## ... consider using the smooth argument
p

## [[1]]
## C
## + -
## 0 1
##
## [[2]]
## C
## A + -
## + NaN 0.25
## - NaN 0.75
##
## [[3]]
## C
## B + -
## + NaN 0.375
## - NaN 0.625
q

## [[1]]
## C
## A + -
## + 0 0.25
## - 0 0.75
##
## [[2]]
## C
## B + -
## + NaN 0.375
## - NaN 0.625
tabProd(p) |> ftable()

## C + -
## A B
## + + NaN 0.09375
## - NaN 0.15625
## - + NaN 0.28125
## - NaN 0.46875
tabProd(q) |> ftable()

## C + -
## A B
## + + NaN 0.09375
## - NaN 0.15625
## - + NaN 0.28125
## - NaN 0.46875
eps <- 0.01
bn01e <- grain(g1, data=tab0, smooth=eps)
bn02e <- grain(g2, data=tab0, smooth=eps)

```



```
j1 <- querygrain(bn01e, type="joint")
j2 <- querygrain(bn02e, type="joint")
j1
```

```
## , , B = +
##
##      A
## C      +      -
## + 0.0006219 0.0006219
## - 0.0941379 0.2805496
##
## , , B = -
##
##      A
## C      +      -
## + 0.0006219 0.0006219
## - 0.1564809 0.4663440
```

```
j2
```

```
## , , B = +
##
##      C
## A      +      -
## + 0.0006219 0.09383
## - 0.0006219 0.28055
##
## , , B = -
##
##      C
## A      +      -
## + 0.0006219 0.1562
## - 0.0006219 0.4670
```

```
d <- j1 %a-% j2
max(abs(d))
```

```
## [1] 0.0006182
```

1.10.2 Building a network from an unknown network

This is a more complex task, sometimes called structural learning. The following code illustrates how to build a network from data. The data is simulated data from the chest clinic example.

We illustrate two approaches: 1) based on the stepwise algorithm in the `gRim` package and 2) based on the hill climbing algorithm in the `bnlearn` package.

```
dat <- gRbase::chestSim10000
```

```
## Using gRim and stepwise selection
sat_model <- gRim::dmod(~.^., data=dat)
mm1 <- stepwise(sat_model, criterion="aic", type="decomposable")
bn1 <- grain(mm1$modelinfo$ug, data=dat)
```

```
## NAs found in clique potential(s) for clique(s):
## c("dysp", "bronc", "tub", "either"), c("asia", "either", "tub", "dysp")
## ... consider using the smooth argument
```

```
bn1 <- grain(mm1$modelinfo$ug, data=dat, smooth=0.01)
## bn1 <- gRim::as.grain(mm1) ## DO WE WANT THIS???
```

```
## Using bnlearn and hill climbing
```

```

sat_graph <- bnlearn::random.graph(names(dat), prob = 1) # complete graph
mm2 <- bnlearn::hc(dat, start=sat_graph)
bn2 <- bnlearn::as.grain(bnlearn::bn.fit(mm2, dat))

```

An important detail is that gRim works with decomposable undirected graphical models (see Figure @ref(fig:bn1)) while bnlearn works with dags (see Figure @ref(fig:bn2), left). However, even for models specified as dags, all computational structures are based on a corresponding undirected graph (see Figure @ref(fig:bn2), left). For details, see the references.

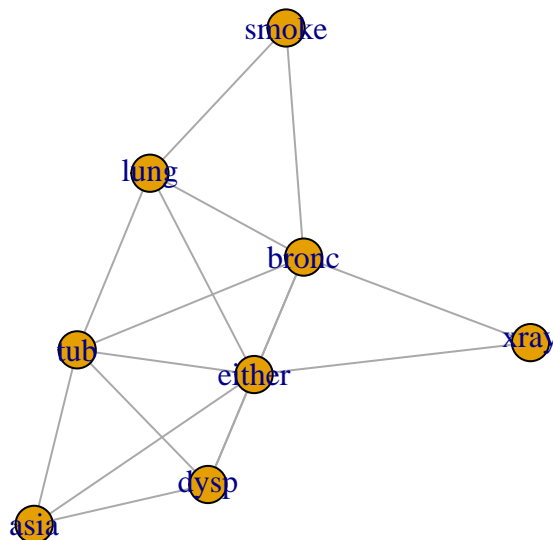


Figure 2: XXXX.

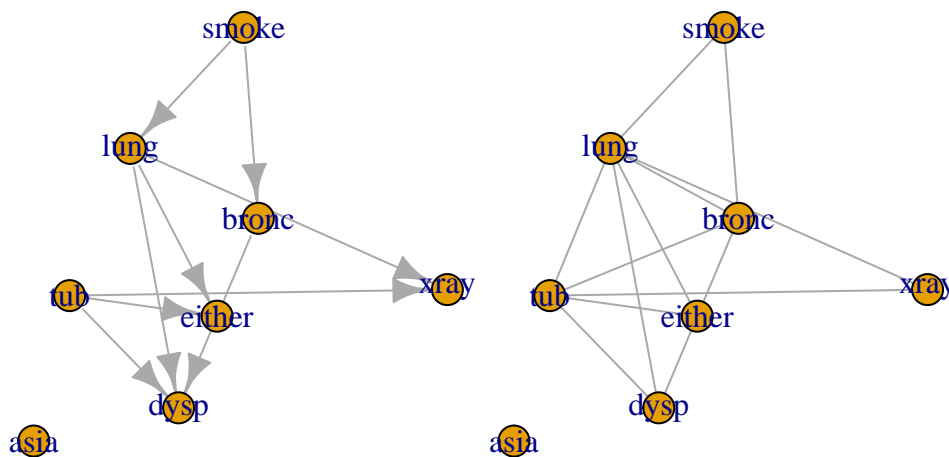


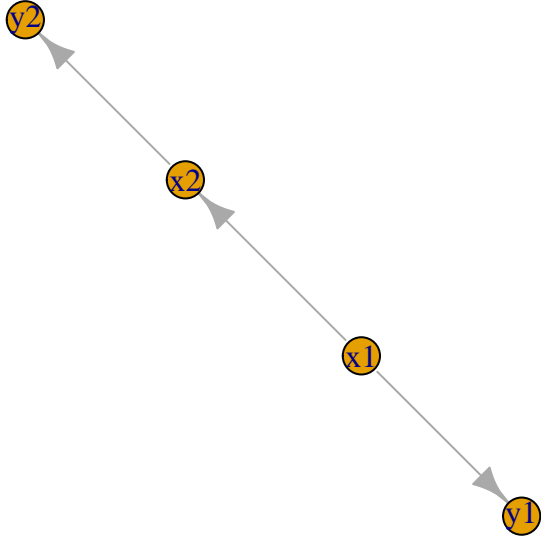
Figure 3: XXXX.

1.11 Extending networks to include other types of variables

gRain only handles discrete variables with a finite state space, but using likelihood evidence it is possible to work with networks with both discrete and continuous variables (or other types of variables). This is possible only when the networks have a specific structure. This is possible when no discrete variable has non-discrete parents.

Take a simple example: Form a network for variables $x = (x_1, x_2)$. Conceptually augment this network with additional variables $y = (y_1, y_2)$ where $y_1|x_1 = k \sim N(\mu_k, v)$ and $y_2|x_2 = k \sim Poi(l_k)$ for $k = 1, 2$. Also we make the assumption that y_1 and y_2 are independent given $x = (x_1, x_2)$. This gives the DAG below:

```
par(mar=c(0,0,0,0))
plot(dag(~y1:x1 + x2:x1 + y2:x2))
```



A network for x can be constructed as:

```
u <- list(x1=yn, x2=yn)
x1 <- cpt(~x1, values=c(1, 3), levels=u)
x2 <- cpt(~x2|x1, values=c(1, 3, 3, 1), levels=u)
bn <- grain(compile_cpt(x1, x2))
querygrain(bn, simplify=TRUE)
```

```
##      yes    no
## x1 0.250 0.750
## x2 0.625 0.375
```

The augmentation of $y|x$ can go along these lines: The parameters describing $y|x$ are set to be:

```
s <- 2
mu <- c(mu1=2, mu2=5)
lambda <- c(lambda1=1, lambda2=7)
```

Suppose we observe $y_1 = y_1^*$. Then

$$p(x|y_1 = y_1^*) \propto p(x_1)p(x_2|x_1)p(y_1 = y_1^*|x_1) = p(x_1)p(x_2|x_1)L_1(x_1) \quad (6)$$

where $L_1(x_1)$ denotes the likelihood. In a network setting this corresponds to changing $p(x_1)$ as

$$p(x_1) \leftarrow p(x_1)L_1(x_1) \quad (7)$$

and then carry on with propagation. This can be achieved in different ways.

The likelihood is:

```
y1_obs <- 14 # Observed value for y1_obs
lik1 <- dnorm(y1_obs, mean=mu, sd=s)
lik1
```

```
##      mu1      mu2
## 3.038e-09 7.992e-06
```

One is by setting the likelihood as evidence. An alternative is to explicitly modify the CPT which specifies $p(x_1)$:

```
querygrain(bn, simplify=TRUE, exclude=FALSE)
```

```
##      yes    no
## x1 0.250 0.750
## x2 0.625 0.375
```

```
bn1 <- setEvidence(bn, evidence=list(x1=lik1))
querygrain(bn1, simplify=TRUE, exclude=FALSE)
```

```
##      yes    no
## x1 0.0001267 0.9999
## x2 0.7499367 0.2501
```

```
x1_upd <- getgrain(bn, "cptlist")$x1 * lik1
bn2 <- replace_cpt(bn, list(x1=x1_upd))
querygrain(bn2, simplify=TRUE)
```

```
##      yes    no
## x1 0.0001267 0.9999
## x2 0.7499367 0.2501
```

A final remark: The conditional distribution of y_1 is normal, but the unconditional distribution is a mixture of normals. Likewise, the conditional distribution of y_2 is poisson, but the unconditional distribution is a mixture of two poisson distributions. Evidence on, say y_1 changes the belief in x_1 and x_2 and this in turn changes the distribution of y_2 (evidence changes the mixture weights.)

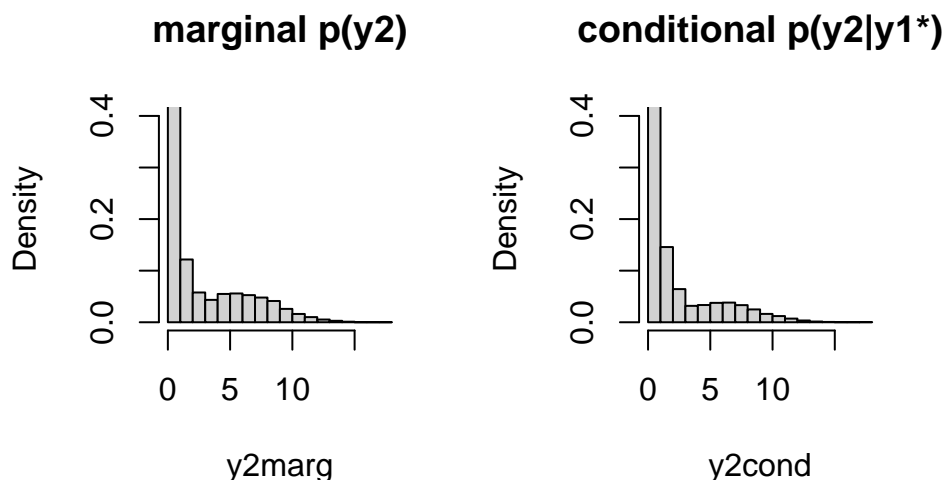
```
nsim <- 10000
xsim_marg <- simulate(bn, nsim, seed=2022)
xsim_cond <- simulate(bn1, nsim, seed=2022)
y2marg <- rpois(n=nsim, lambda=lambda[xsim_marg$x2])
y2cond <- rpois(n=nsim, lambda=lambda[xsim_cond$x2])
summary(y2marg)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.00    1.00    2.00    3.24    6.00   18.00
```

```
summary(y2cond)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.0    0.0    1.0    2.5    3.0   18.0
```

```
par(mfrow=c(1,2))
y2marg |> hist(prob=T, ylim=c(0, .4), breaks=20, main="marginal p(y2)")
y2cond |> hist(prob=T, ylim=c(0, .4), breaks=20, main="conditional p(y2|y1*)")
```



1.12 Brute force computations and why they fail

The gRain package makes computations as those outlined above in a very efficient way; please see the references. However, it is in this small example also possible to make the computations directly: We can construct the joint distribution (an array with $2^8 = 256$ entries) directly as:

```
joint <- tabProd(chest_cpt)
dim(joint)
```

```
## [1] 2 2 2 2 2 2 2 2
```

```
joint |> as.data.frame.table() |> head()
```

```
##   xray smoke asia lung tub dysp bronc either      Freq
## 1  yes   yes  yes  yes yes  yes   yes   yes 1.323e-05
## 2   no   yes  yes  yes yes  yes   yes   yes 2.700e-07
## 3  yes   no   yes  yes yes  yes   yes   yes 6.615e-07
## 4   no   no   yes  yes yes  yes   yes   yes 1.350e-08
## 5  yes   yes   no  yes yes  yes   yes   yes 2.620e-04
## 6   no   yes   no  yes yes  yes   yes   yes 5.346e-06
```

This will clearly fail even moderate size problems: For example, a model with 80 nodes each with 10 levels will give a joint state space with 10^{80} states; that is about the number of atoms in the universe. Similarly, 265 binary variables will result in a joint state space of about the same size. Yet, gRain has been used successfully in models with tens of thousand variables. The “trick” in gRain is to make all computations without ever forming the joint distribution.

However, we can do all the computations by brute force methods as we will illustrate here:

Marginal distributions are

```
tabMarg(joint, "lung")
```

```
## lung
##   yes   no
## 0.055 0.945
```

```
tabMarg(joint, "bronc")
```

```
## bronc
##   yes   no
## 0.45 0.55
```

Conditioning on evidence can be done in different ways: The conditional density is a 6-way slice of the original 8-way joint distribution:

```
asia_dysp
```

```
## $asia
## [1] "yes"
##
## $dysp
## [1] "yes"
```

```
cond1 <- tabSlice(joint, slice=asia_dysp)
cond1 <- cond1 / sum(cond1)
dim(cond1)
```

```
## [1] 2 2 2 2 2 2
```

```
tabMarg(cond1, "lung")
```

```
## lung
##   yes   no
## 0.09953 0.90047
```

```
tabMarg(cond1, "bronc")
```

```
## bronc
##      yes      no
## 0.8114 0.1886
```

Alternatively, multiply all entries not consistent by zero and all other entries by one and then marginalize:

```
cond2 <- tabSliceMult(joint, slice=asia_dysp)
cond2 <- cond2 / sum(cond2)
dim(cond2)
```

```
## [1] 2 2 2 2 2 2 2 2
```

```
tabMarg(cond2, "lung")
```

```
## lung
##      yes      no
## 0.09953 0.90047
```

```
tabMarg(cond2, "bronc")
```

```
## bronc
##      yes      no
## 0.8114 0.1886
```

Højsgaard, Søren. 2012. "Graphical Independence Networks with the gRain Package for R." *Journal of Statistical Software* 46 (10): 1–26. <http://www.jstatsoft.org/v46/i10/>.

Højsgaard, Søren, David Edwards, and Steffen L. Lauritzen. 2012. *Graphical Models with r*. Springer.

Lauritzen, Steffen L., and David Spiegelhalter. 1988. "Local Computations with Probabilities on Graphical Structures and Their Application to Expert Systems." *J. Roy. Stat. Soc. Ser. B* 50 (2): 157–224.